

TECHNICAL ADVANCE

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Soluble Fas might serve as a diagnostic tool for gastric adenocarcinoma

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Abstract

Background: Fas (Apo-1/CD95) and its specific ligand (FasL) are key elements in apoptosis. They have been studied in different malignancies but there are few published studies about the soluble forms of these markers (i.e. sFas/sFasL) in gastric cancer. We have compared the serum levels of sFas/sFasL in gastric adenocarcinoma patients and cases with pre-neoplastic lesions as potential markers for early diagnosis, and investigated their relation with clinicopathological characteristics.

Methods: Fifty-nine newly-diagnosed cases of gastric adenocarcinoma who had undergone gastrectomy, along with 62 endoscopically- and histologically-confirmed non-cancer individuals were enrolled in this study. sFas/sFasL serum levels were detected by Enzyme Linked Immunosurbent Assay.

Results: Mean serum sFas level was significantly higher in gastric cancer patients than in control group (305.97 \pm 63.71 (pg/ml) vs. 92.98 \pm 4.95 (pg/ml), P < 0.001); while the mean serum level of sFasL was lower in patients with gastric adenocarcinoma (0.138 \pm 0.04 (pg/ml) vs. 0.150 \pm 0.02 (pg/ml), P < 0.001). Mean serum levels of sFas/sFasL were significantly different in both intestinal/diffuse and cardiac/non-cardiac subtypes when compared to the control group (P < 0.001). There was an increase in the serum level of sFas from the first steps of pre-neoplastic lesions to gastric adenocarcinoma (P < 0.001). Patients who had no lymph node involvement (N_0) showed significantly higher serum levels of sFas compared to others (P = 0.044).

Conclusions: Production of sFas may play a critical role in the carcinogenesis of intestinal-type gastric cancer. sFas serum level may serve as a non-invasive tool for early diagnosis of gastric cancer.

Background

Gastric cancer is the second leading cause of cancerrelated mortality worldwide with a great geographic variation[1]. In Iran, it is the second most common cancer in males and the forth in females. It has been estimated that gastric cancer is the most common cause of cancer-associated mortality in Iranian population[2].

Although gastric cancer has a poor prognosis with a five-year survival rate of 25% in the United States[3], there is no standard biomarker for early diagnosis and no consensus on screening programs. To date, some guidelines have been proposed for early diagnosis[1] and new molecular markers and therapeutic strategies are required to design effective diagnostic and therapeutic protocols.

Derailment of apoptosis plays an important role in the development, growth and resistance of malignant tumors, and also influences the prognosis[4]. As a member of TNF-family receptors, Fas (Apo-1/CD95) is a cell surface protein that can induce apoptosis through its cytosolic tail after binding to its specific ligand, Fas Ligand[5]. Fas and Fas Ligand (FasL) are crucial in immune system homeostasis[6,7]. FasL is also a major weapon for cytolytic T cells to induce apoptosis in tumor cells[4]. Consequently, a decrease in Fas expression in the membrane of tumor cells can protect them from this lethal influence of FasL. FasL detection on the cell surface of some tumor cells proposed the hypothesis that these

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cells can escape immune attack through induction of apoptosis in tumor infiltrating lymphocytes[8,9]. The *Fas* gene produces two protein isoforms through alternative mRNA splicing: the full-length Fas which contains a trans-membrane domain, and the soluble form of Fas (sFas), which lacks this domain. Soluble Fas acts as a decoy in the extra-cellular environment and binds FasL[10,11]. Therefore, there is an immune privilege for tumor cells by secretion of sFas as an inhibitor of apoptosis

Fas/FasL system has been investigated in a large variety of neoplasms[12-15]. However, few studies have been reported about gastric cancer to date. Previous studies indicate that gastric carcinomas express FasL at a higher level, while lower level of Fas expression leads to evade the killing effects of host immune system[16-18]. There are some conflicting reports about the correlations of Fas/FasL expression--studied by immunohistochemistry (IHC) method--and tumor size, depth of invasion, metastasis, differentiation and Lauren's classification of gastric tumors [18,19]. Few studies have been published about the serum level of sFas/sFasL in gastric cancer, with discrepancies in their results[20-23].

This study was conducted to assess the serum level of sFas/sFasL in gastric adenocarcinoma and non-tumoral lesions, to find their possible role in early diagnosis and their correlations with clinicopathological features of this malignancy.

Methods

Sample collection

Study included fifty-nine patients with newly-diagnosed, histologically-confirmed gastric adenocarcinoma, who were admitted to the department of surgery in Omid Oncology Hospital, Mashhad, Iran; between February 2006 and June 2008. Patients with unresectable tumors, history of previous chemotherapy, radiation therapy, or gastric surgery were excluded. Sixty-two individuals were enrolled in the control group. They had all undergone esophagogastrodeodenoscopy due to upper gastrointestinal complaints in the endoscopy unit, Imam Reza University Hospital, Mashhad, Iran; and were proven to have no endoscopic and histological evidence of gastric tumor.

The study protocol was approved by the Research Ethics Committee in Mashhad University of Medical Sciences. A written informed consent was obtained from each individual. Demographic characteristics (e.g. ethnicity, age, and gender) and clinical data including symptoms, medications, and potential risk factors were obtained via a questionnaire filled by trained personnel. These risk factors included history of tobacco consumption and family history of gastric cancer.

According to updated Sydney System[24], five biopsies were obtained from each individual in the control group

for histological examination: two from the antrum, two from the corpus, and one from the incisura angularis. Another antral biopsy was obtained for the detection of H. pylori via a commercially available Rapid Urease Test (Chemenzyme Co., Iran). Biopsy samples were fixed in 10% buffered formalin. After routine tissue processing, they were examined for the presence of five pathologic variables including density of H. pylori, intensity of neutrophilic and mononuclear inflammation, atrophy, intestinal metaplasia, and dysplasia. We divided the control group into three subgroups according to the pathologic pattern: mild gastritis without H. pylori infection (named as near-normal mucosa), chronic gastritis with H. pylori infection (chronic active gastritis), and precancerous lesions (including chronic atrophic gastritis, gastric atrophy, intestinal metaplasia, and dysplasia). Giemsa staining was applied in suspicious cases for better evaluation of H. pylori.

In the cancer group, the histological diagnosis was based on morphological examination of the samples that were routinely processed and stained with hematoxylin and eosin method. According to Lauren's criteria, tumors were classified as intestinal and diffuse type. Tumor grade and surgical stage were determined as well.

Enzyme Linked Immunosurbent Assay (ELISA)

A 3-ml sample of venous blood was collected from each participant before endoscopy or one day before surgery, in control and cancer groups, respectively. Immediately after blood sampling, serum was obtained by centrifugation at 2000 r/min for 15 min at 4°C and stored at -20°C until subsequent assay. The titres of *H. pylori* IgG antibody were measured via commercial ELISA (Padtan Elm Co., Ltd. Iran), according to the manufacturer's instructions. Serum levels of sFas and sFasL were assessed using human ELISA kits (Bender MedSystems, GmbH, Vienna, Austria) according to the manufacturer's instructions.

Statistical analysis

Chi-square test with 95% confidence interval was performed for comparing the variables. As serum levels of sFas and sFasL did not have a normal distribution, we used Kruskal-Wallis and Mann-Whitney U tests to compare the mean serum levels of sFas and sFasL in different groups. Data have been shown as mean \pm SEM (standard error of mean) in ures and tables. The statistical analyses were performed using the SPSS 16.0 statistical package (SPSS, Inc, Chicago, IL, USA).

Results

Forty-four out of 59 patients with gastric adenocarcinoma were male (M/F ratio: 2.93). The median age was 62 years (ranging from 39 to 79, mean: 60.25 ± 10). Of 62 individuals in the non-tumoral group, 32 were males and

30 were females with a median age of 47 years (ranging from 20 to 77, mean: 47.32 ± 16). Poor economic status, lower level of education and tobacco consumption were significantly more prevalent in cancer group (p < 0.05). Most of our patients were diagnosed as intestinal type gastric cancer (45/59), of which 10, 15 and 5 were well, moderately and poorly differentiated carcinomas, respectively, and in 5 cases, the grade of differentiation was unknown. Among 62 non-cancer individuals 11 were categorized as near-normal mucosa, 35 as chronic active gastritis and in 16 cases, precancerous lesions were seen in the stomach. Based on laboratory exams, 73.6% of our cancer patients had a positive history of H. pylori infection, while in the control group this positive history was found in 82.3%.

Mean serum levels of sFas/sFasL based on descriptive characteristics of the non-tumoral and tumoral groups are summarized in table 1 and table 2. There was a positive correlation between tobacco smoking and mean serum level of sFas, but not sFasL, among non-tumoral

group (P = 0.041). However, in the tumoral group, no association was observed between smoking and mean serum level of sFas/sFasL (P = 0.06 for sFas). There was no significant difference in serum levels of sFas/sFasL between cases with or without history of H. pylori infection in either tumoral or non-tumoral groups.

The mean serum level of sFas was significantly higher in gastric cancer patients than control group (P < 0.001), while the mean serum level of sFasL was lower in patients with gastric adenocarcinoma (P < 0.001). After grouping the patients by histological type (intestinal/diffuse) and tumor location (cardia/non-cardia), there were statistically significant differences in serum sFas/sFasL level in all of the subgroups versus non-tumoral group (Table 3).

An increasing gradient for mean serum level of sFas was observed, from normal mucosa toward gastric cancer (Kruskal-Wallis test, P < 0.001). Analysis showed a significant difference between serum level of sFas in each nontumoral subgroup and cancer group (Figure 1), (tumoral

Table 1: Serum levels of sFas/sFasL based on descriptive characteristics of non-tumoral group

	Non-Tumoral Group						
	sFas (pg/ml)			sFasL (pg/ml)			
_	N*	Mean ± SE.M†	P Value	N*	Mean ± SE.M†	P Value	
Gender							
Male	30	103.21 ± 7.54	0.023	32	0.13 ± 0.02	0.09	
Female	29	82.40 ± 5.88		30	0.17 ± 0.03		
Economic Status\							
Poor	15	85.21 ± 11.66	0.82	16	0.10 ± 0.03	0.17	
Moderate to Good	43	95.07 ± 5.45		45	0.16 ± 0.02		
Educational Status							
Under high school Diploma	41	97.94 ± 6.44	0.25	43	0.16 ± 0.03	0.91	
High school Diploma & above	16	81.42 ± 6.91		17	0.13 ± 0.02		
Tobacco consumption							
Negative	51	88.83 ± 5.03	0.041	53	0.16 ± 0.02	0.11	
Positive	8	119.46 ± 15.27		9	0.11 ± 0.01		
Opium addiction							
Negative	46	86.81 ± 5.20	0.015	47	0.16 ± 0.02	0.52	
Positive	13	114.82 ± 11.31		15	0.12 ± 0.02		
History of H. pylori infection‡							
Negative	49	76.17 ± 9.76	0.12	51	0.14 ± 0.04	0.27	
Positive	10	97.74 ± 5.58		11	0.15 ± 0.02		

^{*} N: Number of samples

[†] Mean ± SE.M: Mean Serum Range ± Standard Error of Mean

[‡] History of H. pylori infection: Based on laboratory data

Table 2: Serum levels of sFas/sFasL based on descriptive characteristics of patients with gastric adenocarcinoma

	Gastric Adenocarcinoma Group						
	sFas (pg/ml)			sFasL (pg/ml)			
	N*	Mean ± SE.M†	P Value	N*	Mean ± SE.M†	P Value	
Gender							
Male	41	348.89 ± 85.04	0.55	44	0.16 ± 0.06	0.28	
Female	15	188.63 ± 4.54		15	0.06 ± 0.04		
Economic Status							
Poor	24	222.94 ± 45.98	0.93	26	0.05 ± 0.02	0.006	
Moderate to Good	20	279.68 ± 104.11		21	0.20 ± 0.11		
Educational Status							
Under high school Diploma	38	262.75 ± 61.03	0.33	41	0.08 ± 0.03	0.018	
High school Diploma & above	4	129.93 ± 35.48		4	0.60 ± 0.54		
Tobacco consumption							
Negative	28	234.81 ± 76.62	0.06	29	0.07 ± 0.03	0.34	
Positive	15	281.3 ± 65.87		17	0.22 ± 0.13		
Opium addiction							
Negative	46	282.55 ± 64.73	0.37	49	0.15 ± 0.05	0.47	
Positive	10	413.67 ± 202.46		10	0.10 ± 0.06		
History of <i>H. pylori</i> infection [‡]							
Negative	34	390.46 ± 145.66	0.64	37	0.10 ± 0.04	0.13	
Positive	15	182.09 ± 21.08		15	0.14 ± 0.07		

^{*} N: Number of samples

vs. normal: P < 0.001; tumoral vs. chronic active gastritis: P < 0.001; tumoral vs. precancerous lesions: P = 0.009).

Serum level of sFas/sFasL and their correlation with clinicopathological features of the tumors are represented in Table 4. Although serum level of sFasL was significantly lower in cardiac type of tumor versus non-cardiac type (P = 0.005), serum level of sFas was not significantly associated to tumor location (cardia vs. non-cardia). There was no correlation between serum levels of sFas/sFasL and different histopathologic subtypes of adenocarcinoma (intestinal vs. diffuse), tumor grade of differentiation and stage of tumor. Patients with no lymph node metastasis (N_0) had significantly higher levels of sFas than those with lymph node involvement (N_{1-3}) (P = 0.044).

Discussion

Gastric cancer is the second leading cause of cancerrelated deaths worldwide. It is diagnosed in advanced stages in the majority of cases and no efficient therapeutic modality has been suggested to overcome the problem of treatment resistance yet. Gastric adenocarcinoma has a complex network of molecular alterations along its carcinogenesis pathway. Despite numerous studies focused on this issue, many crucial questions still remain to be clarified. Discovery of these molecular changes can be translated into efficient diagnostic and therapeutic modalities and be employed for targeting the cancer like in some other malignancies[25,26]. Apoptosis-regulating genes play a critical role in carcinogenesis. Fas/FasL system exerts a central role in the apoptosis process and its alterations are noticeable in gastric adenocarcinoma. Although there are some studies indicating that gastric carcinomas express higher levels of FasL and lower levels of Fas to evade the killing effects of host immune system[16-18], there are only few reports addressing their soluble forms.

While Yatsuya *et al* reported significant difference in serum concentration of sFas only between female gastric cancer patients and controls[23], Liang *et al* represented a significantly higher serum level of sFas in all patients

 $^{^{\}dagger}$ Mean \pm SE.M: Mean Serum Range \pm Standard Error of Mean

[‡] History of H. pylori infection: Based on laboratory data

Table 3: Comparison of serum levels of sFas and sFasL in different subgroups of gastric adenocarcinoma with non-tumoral group

	sFas (pg/ml)				sFasL (pg/ml)			
	N*	Mean ± SE.M†	P Value	N*	Mean ± SE.M†	P Value		
Gastric Cancer Group			(vs. Non-Tumoral)			(vs. Non-Tumoral)		
Overall	56	305.97 ± 63.71	< 0.001	59	0.138 ± 0.04	< 0.001		
Intestinal	43	293.06 ± 73.88	< 0.001	45	0.147 ± 0.05	< 0.001		
Diffuse	13	348.61 ± 129.43	< 0.001	14	0.109 ± 0.05	< 0.001		
Cardia	18	242.72 ± 62.78	0.001	18	0.026 ± 0.01	< 0.001		
Non-cardia	37	358.92 ± 99.61	< 0.001	40	0.206 ± 0.07	< 0.001		
Non-Tumoral Group	59	92.98 ± 4.95		62	0.150 ± 0.02			

^{*} N: Number of samples

with gastric cancer compared to non-tumoral individuals[21], similar to our results. To explain the observation of decrease in Fas and increase in serum level of sFas in gastric cancer[17,21], we hypothesize that translational processing of the Fas gene in gastric tumoral cells may be deranged leading to the production of mostly soluble, rather than membranous, Fas. Possibly, mRNA splicing phase alternates toward producing higher levels of sFas with lower molecular weight rather than full-length (membranous) Fas. Thus, we conclude that changes in Fas gene expression may be a part of sequential events in the multistep process of gastric cancer development.

To confirm this idea, a group of non-tumoral cases, consisting of various pathologic lesions of non-precancerous and precancerous lesions from different levels of multistep carcinogenesis pathway, were evaluated for serum levels of sFas and sFasL. The findings indicated an increasing gradient in the level of sFas from normal through tumoral epithelium. Analysis showed a significant difference between sFas level in each noncancerous subgroup and cancer group; in line with the findings of Li et al[27]. With the application of IHC and western-blot hybridization methods, they reported increasing frequencies of Fas expression in progression from non-cancerous to cancerous mucosa (6.3% in normal mucosa, 60% in atrophic gastritis, 75% in intestinal metaplasia, 100% in grades 2 and 3 dysplasia and gastric adenocarcinoma). They found that soluble Fas (30 KD), but not the membrane type (43 KD), was predominantly expressed in the Fas-positive cases[27]. By quantitatively measuring the increased levels of sFas in serum, we suggest that production of sFas is a crucial event in gastric carcinogenesis. In addition, the significant difference between the serum levels in gastric cancer patients and precancerous group, and also the increases along the carcinogenesis pathway may introduce sFas as a useful, cost-effective, and non-invasive biomarker for early detection of gastric cancer. Further studies with larger sample size are required to establish a precise cut-off point for that purpose. Tamakoshi A *et al* performed a nested case-control study within a large-scale prospective study and suggested that serum sFas has a possibility to detect people at high risk for cancer (regardless of cancer type) prior to diagnosis[15].

We found a significantly higher serum level of sFasL in patients with non-cardiac type of gastric cancer versus those with cardiac type (P = 0.005). To our knowledge, there is no published study addressing the serum levels of sFas/sFasL in cardiac and non-cardiac types of gastric cancer. Our results may confirm the differences in the carcinogenesis pathway and molecular alterations of these subgroups of gastric tumors. Further studies are required to clarify the role of sFas/sFasL in gastric carcinogenesis in cardiac versus non-cardiac tumors.

Some studies reported elevated concentrations of serum sFasL in patients with various types of malignancies and concluded that sFasL may be derived from cancer cells as a result of high expression of FasL gene[28-30]. Others have shown lower levels of sFasL in cancer patients compared to controls, suggesting that serum sFasL is possibly consumed by binding to Fas expressed on activated circulating CD8+T lymphocytes[31,32]. In gastric cancer, there are few reports with controversial findings to date. Yoshikawa et al showed lower level of sFasL in serum of patients with gastric cancer than normal controls[32], while significantly higher level of sFasL

[†] Mean ± SE.M: Mean Serum Range ± Standard Error of Mean

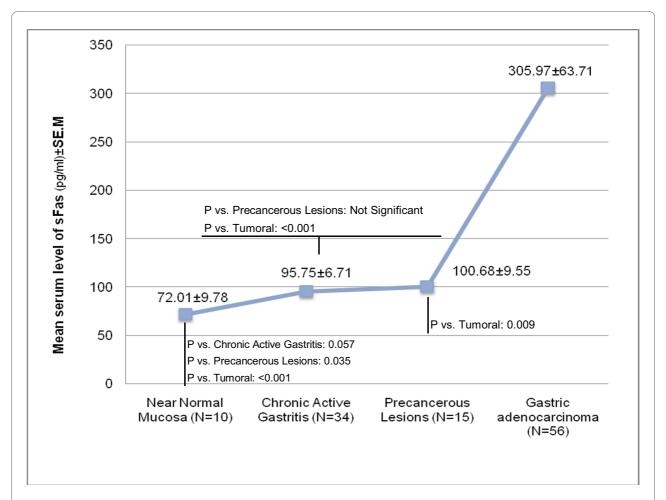


Figure 1 Mean serum level of sFas (pg/ml) in intestinal-type gastric adenocarcinoma patients and different non-tumoral subgroups. The graph represents an increasing gradient for the mean serum level of sFas from normal mucosa toward gastric cancer (Kruskal-Wallis test, P < 0.001). Significant difference was observed between serum levels of sFas in each non-tumoral subgroup and cancer group.

in serum was reported by Ichikura et al, only in older male patients (over 50 years old)[20]. In contrast, Tsutsumi et al found not statistically significant differences between serum level of sFasL in cancer patients compared to normal individuals[22]. In the studied population, serum level of sFasL was significantly lower in tumoral than non-tumoral group. As the serum level of sFasL could be influenced by both tumor cells production and immune cells consumption of sFasL, the discrepancies in the results may be explained by differences in immune responses of patients. Varieties in clinicopathological features and diversities in socio-demographic characteristics and ethnic background of the studied populations may be another reason for controversial results. Further validation sets focusing on cell expression of FasL, serum level of sFasL and concurrent evaluation of different aspects and impacts of immune response could better elucidate the role of FasL/sFasL in gastric carcinogenesis.

There are few reports about the relation between serum level of soluble Fas and gastric cancer behaviour. Liang *et al* found a direct relation between increasing sFas level with advance in the tumor grade and stage [21]. We observed a lower serum level of sFas in patients with lymph node involvement. When tumor involves lymph nodes, antitumor immunity will be provoked [33] which may result in production of more FasL bearing immune cells and subsequently sFas may be consumed more following binding and neutralizing these Fas Ligands.

Few controversial studies regarding the effects of consuming to bacco on Fas signalling pathways have been published. Some have shown that to bacco increases apoptosis through Fas signalling pathway [34,35], while others reported the anti-apoptotic effects of to bacco on the pathway [36,37]. We found higher levels of sFas in to bacco users of the control group (p = 0.041), which is in favour of apoptotic-inducing role of smoking. However, as most of the cases were not to bacco users (53/62), stud-

Table 4: Serum levels of sFas/sFasL and their correlations with clinicopathological features of patients with gastric adenocarcinoma

	sFas (pg/ml)			sFasL (pg/ml)			
-	N*	Mean ± SE.M†	P Value	N*	Mean ± SE.M†	P Value	
Tumor invasion							
T1 & T2	5	228.66 ± 50.29	0.45	5	0.46 ± 0.44	0.93	
T3 & T4	36	227.73 ± 64.32		38	0.09 ± 0.03		
Lymph node involvement							
NO	14	404.86 ± 150.62	0.044	14	0.24 ± 0.16	0.57	
N >0	28	273.00 ± 68.97		30	0.11 ± 0.03		
Stage							
1&11	22	479.83 ± 151.47	0.2	23	0.22 ± 0.10	0.16	
III & IV	32	199.70 ± 28.16		34	0.09 ± 0.03		
Grade							
Well Differentiated	11	392.19 ± 209.08	0.75	11	0.29 ± 0.20	0.92	
Moderatly Differentiated	15	307.88 ± 136.59		15	0.11 ± 0.05		
Poorly Differentiated	4	334.00 ± 232.12		5	0.11 ± 0.08		
Tumor location							
Cardia	18	242.72 ± 62.78	0.53	18	0.03 ± 0.01	0.005	
Non-cardia	37	358.92 ± 99.61		40	0.21 ± 0.07		

^{*} N: Number of samples

ies with larger sample size and greater statistical power are required to confirm this finding.

Conclusions

Fas/FasL system plays a crucial role in gastric carcinogenesis. Assessing the level of sFas in serum, may serve as a biomarker for early diagnosis of gastric cancer. Further studies that investigate both soluble and membranous isoforms of *Fas* gene products may provide valuable information about correlations between serum level and tissue expression of *Fas* gene products, helping in gaining a better understanding of molecular basis of these changes.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

SB-N, HRS, KG, HRR, MJ and JT-A conceived and designed the study, SB-N, KG, carried out the experimental work, SB-N, HH, MJ, OM and MM participated in the classification and characterization of patients, HRS, MTR-M, HH and MM were involved in the recruitment of patients and controls. SB-N, HRS, HRR, MJ, OM and AAA performed the statistical analysis and wrote the manuscript. All authors helped to draft the manuscript and read and approved this final version.

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References

- Dicken BJ, Bigam DL, Cass C, Mackey JR, Joy AA, Hamilton SM: Gastric adenocarcinoma: review and considerations for future directions. Ann Surg 2005, 241(1):27-39.
- Mousavi SM, Gouya MM, Ramazani R, Davanlou M, Hajsadeghi N, Seddighi Z: Cancer incidence and mortality in Iran. Ann Oncol 2009, 20(3):556-563.
- Jemal A, Siegel R, Ward E, Hao Y, Xu J, Thun MJ: Cancer Statistics, 2009. CA Cancer J Clin 2009. 59:225-249.

 $^{^{\}dagger}$ Mean \pm SE.M: Mean Serum Range \pm Standard Error of Mean

- Reed JC: Apoptosis and cancer. In Cancer medicine 6th edition. Edited by: Kufe DW, Pollock RE, Weichselbaum RR, Bast RC Jr. Hamilton: BC Decker; 2003:41-52.
- Locksley RM, Killeen N, Lenardo MJ: The TNF and TNF receptor superfamilies: integrating mammalian biology. Cell 2001, 104(4):487-501.
- Krammer PH: CD95's deadly mission in the immune system. *Nature* 2000, 407(6805):789-795.
- Ju ST, Panka DJ, Cui H, Ettinger R, el-Khatib M, Sherr DH, Stanger BZ, Marshak-Rothstein A: Fas(CD95)/FasL interactions required for programmed cell death after T-cell activation. Nature 1995, 373(6513):444-448.
- Li-Weber M, Krammer PH: Function and regulation of the CD95 (APO-1/ Fas) ligand in the immune system. Semin Immunol 2003, 15(3):145-157.
- Igney FH, Krammer PH: Immune escape of tumors: apoptosis resistance and tumor counterattack. J Leukoc Biol 2002, 71(6):907-920.
- Cascino I, Fiucci G, Papoff G, Ruberti G: Three functional soluble forms of the human apoptosis-inducing Fas molecule are produced by alternative splicing. *J Immunol* 1995, 154(6):2706-2713.
- Cheng J, Zhou T, Liu C, Shapiro JP, Brauer MJ, Kiefer MC, Barr PJ, Mountz JD: Protection from Fas-mediated apoptosis by a soluble form of the Fas molecule. Science 1994, 263(5154):1759-1762.
- 12. Munakata S, Enomoto T, Tsujimoto M, Otsuki Y, Miwa H, Kanno H, Aozasa K: Expressions of Fas ligand and other apoptosis-related genes and their prognostic significance in epithelial ovarian neoplasms. *Br J Cancer* 2000, **82(8)**:1446-1452.
- Bebenek M, Dus D, Kozlak J: Fas expression in primary breast cancer is related to neoplastic infiltration of perilymphatic fat. Adv Med Sci 2008, 53(1):49-53.
- Kamihira S, Yamada Y, Tomonaga M, Sugahara K, Tsuruda K: Discrepant expression of membrane and soluble isoforms of Fas (CD95/APO-1) in adult T-cell leukaemia: soluble Fas isoform is an independent risk factor for prognosis. Br J Haematol 1999, 107(4):851-860.
- Tamakoshi A, Nakachi K, Ito Y, Lin Y, Yagyu K, Kikuchi S, Watanabe Y, Inaba Y, Tajima K: Soluble Fas level and cancer mortality: findings from a nested case-control study within a large-scale prospective study. Int J Cancer 2008, 123(8):1913-1916.
- Bennett MW, O'connell J, O'sullivan GC, Roche D, Brady C, Kelly J, Collins JK, Shanahan F: Expression of Fas ligand by human gastric adenocarcinomas: a potential mechanism of immune escape in stomach cancer. Gut 1999, 44(2):156-162.
- Li Z, Wang Z, Zhao Z, Zhang Y, Ke Y: [Expression of Fas, FasL and IFNgamma in gastric cancer]. Beijing Da Xue Xue Bao 2003, 35(4):386-389.
- 18. Zheng HC, Sun JM, Wei ZL, Yang XF, Zhang YC, Xin Y: Expression of Fas ligand and caspase-3 contributes to formation of immune escape in gastric cancer. *World J Gastroenterol* 2003, **9(7)**:1415-1420.
- Ohno S, Tachibana M, Shibakita M, Dhar DK, Yoshimura H, Kinugasa S, Kubota H, Masunaga R, Nagasue N: Prognostic significance of Fas and Fas ligand system-associated apoptosis in gastric cancer. Ann Surg Oncol 2000, 7(10):750-757.
- Ichikura T, Majima T, Uchida T, Okura E, Ogawa T, Mochizuki H: Plasma soluble Fas ligand concentration: decrease in elderly men and increase in patients with gastric carcinoma. Oncol Rep 2001, 8(2):311-314.
- Liang QL, Pan DC, Yin ZM, Liu GX, Yang Q, Xie JR, Cai LZ, Fu YW: [Clinical value of serum soluble Apo-1/Fas for predicting biological behaviors and prognosis of gastric carcinoma]. Ai Zheng 2002, 21(2):174-176.
- Tsutsumi S, Kuwano H, Shimura T, Morinaga N, Mochiki E, Asao T: Circulating soluble Fas ligand in patients with gastric carcinoma. Cancer 2000, 89(12):2560-2564.
- Yatsuya H, Toyoshima H, Tamakoshi K, Tamakoshi A, Kondo T, Hayakawa N, Sakata K, Kikuchi S, Hoshiyama Y, Fujino Y, Mizoue T, Tokui N, Yoshimura T: Serum levels of insulin-like growth factor I, II, and binding protein 3, transforming growth factor beta-1, soluble fas ligand and superoxide dismutase activity in stomach cancer cases and their controls in the JACC Study. J Epidemiol 2005, 15(Suppl 2):S120-125.
- Dixon MF, Genta RM, Yardley JH, Correa P: Classification and grading of gastritis. The updated Sydney System. International Workshop on the Histopathology of Gastritis, Houston 1994. Am J Surg Pathol 1996, 20(10):1161-1181.
- Bremer E, de Bruyn M, Wajant H, Helfrich W: Targeted cancer immunotherapy using ligands of the tumor necrosis factor superfamily. Curr Drug Targets 2009, 10(2):94-103.

- Rikhof B, van der Graaf WT, Meijer C, Le PT, Meersma GJ, de Jong S, Fletcher JA, Suurmeijer AJ: Abundant Fas expression by gastrointestinal stromal tumours may serve as a therapeutic target for MegaFasL. Br J Cancer 2008, 99(10):1600-1606.
- 27. Li H, Liu N, Guo L, Li JW, Liu J: Frequent expression of soluble Fas and Fas ligand in Chinese stomach cancer and its preneoplastic lesions. *Int J Mol Med* 2000, **5(5)**:473-476.
- Bellone G, Smirne C, Carbone A, Mareschi K, Dughera L, Farina EC, Alabiso O, Valente G, Emanuelli G, Rodeck U: Production and pro-apoptotic activity of soluble CD95 ligand in pancreatic carcinoma. clin Cancer Res 2000. 6(6):2448-2455.
- Mizutani Y, Hongo F, Sato N, Ogawa O, Yoshida O, Miki T: Significance of serum soluble Fas ligand in patients with bladder carcinoma. *Cancer* 2001, 92(2):287-293.
- Tanaka M, Suda T, Haze K, Nakamura N, Sato K, Kimura F, Motoyoshi K, Mizuki M, Tagawa S, Ohga S, Hatake K, Drummond AH, Nagata S: Fas ligand in human serum. Nat Med 1996, 2(3):317-322.
- Hoffmann TK, Dworacki G, Tsukihiro T, Meidenbauer N, Gooding W, Johnson JT, Whiteside TL: Spontaneous apoptosis of circulating T lymphocytes in patients with head and neck cancer and its clinical importance. Clin Cancer Res 2002, 8(8):2553-2562.
- Yoshikawa T, Saito H, Osaki T, Matsumoto S, Tsujitani S, Ikeguchi M: Elevated Fas expression is related to increased apoptosis of circulating CD8+ T cell in patients with gastric cancer. J Surg Res 2008, 148(2):143-151.
- 33. Kim R, Emi M, Tanabe K, Arihiro K: Immunobiology of the sentinel lymph node and its potential role for antitumour immunity. *Lancet Oncol* 2006, **7(12)**:1006-1016.
- Kuo WH, Chen JH, Lin HH, Chen BC, Hsu JD, Wang CJ: Induction of apoptosis in the lung tissue from rats exposed to cigarette smoke involves p38/JNK MAPK pathway. Chem Biol Interact 2005, 155(1-2):31-42
- Park JW, Kim HP, Lee SJ, Wang X, Wang Y, Ifedigbo E, Watkins SC, Ohba M, Ryter SW, Vyas YM, Choi AM: Protein kinase C alpha and zeta differentially regulate death-inducing signaling complex formation in cigarette smoke extract-induced apoptosis. J Immunol 2008, 180(7):4668-4678.
- 36. Imirzalioglu P, Uckan S, Alaaddinoglu EE, Haberal A, Uckan D: Cigarette smoking and apoptosis. *J Periodontol* 2005, **76(5)**:737-739.
- 37. Sarafian TA, Tashkin DP, Roth MD: Marijuana smoke and Delta(9)-tetrahydrocannabinol promote necrotic cell death but inhibit Fasmediated apoptosis. *Toxicol Appl Pharmacol* 2001, **174(3)**:264-272.

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