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# Retinol-binding protein type 1 expression predicts poor prognosis in head and neck squamous cell carcinoma

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# **Abstract**

**Background** Head and neck squamous cell carcinoma (HNSCC) is the sixth most prevalent malignancy worldwide, with high incidence and poor survival rates. RBP1 is highly expressed in several kinds of cancer and plays a potential prognostic factor. However, the relationship between RBP1 and HNSCC were analyzed based on The Cancer Genome Atlas (TCGA) database.

**Materials and methods** RBP1 expression and clinical information were obtained from the Cancer Genome Atlas (TCGA) database. Tumor tissue and adjacent normal tissue of 6 HNSCC patients were collected to analyze the RBP1 mRNA expression level by quantitative PCR. Cox regression analysis was used to evaluate the prognostic values of RBP1 and clinical data in HNSCC. A nomogram was also established to predict the impact of RBP1 on prognosis based on Cox multivariate results. The methylation level of RBP1 in HNSC and its prognosis were analyzed in UALACN and MethSurv. Finally, the potential biological functions of RBP1 were investigated using gene set enrichment analysis (GSEA) and single sample GSEA (ssGSEA).

**Results** The mRNA expression levels of RBP1 were highly expressed in HNSCC tissue. The Cox analyses demonstrate that highly-expressed RBP1 is an independent prognosis marker(*P*<0.05). ROC curve analysis showed that performances of RBP1 (area under the ROC curve: 0.887, sensitivity: 84.1%, specificity: 79.9%). The methylation was increased in HNSCC patients compared with normal subjects(*P*<0.05) and was associated with better prognosis at sites cg06208339, cg12298268, cg12497564, cg15288618, cg20532370, cg23448348. Additionally, RBP1 expression is mildly associated with immune cell infiltration and immunological checkpoints.

**Conclusion** RBP1 is overexpressed and associated with poor patient prognosis in head and neck squamous cell carcinoma.

**Keywords** RBP1, Bioinformatic analysis, Prognosis biomarker, Head and neck squamous cell carcinoma

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# **Introduction**

Head and neck tumor is the sixth most common malignancy in the world, with 830 thousand new cases diagnosed annually. Head and neck squamous cell carcinoma (HNSCC) comprises 90% of head and neck tumors [\[1](#page-11-0), [2\]](#page-11-1). For the past few years, advances in treatment, such as surgery, radiotherapy, and chemotherapy, have contributed to certain improvement in clinical outcomes [[3\]](#page-11-2). However, the 5-year survival rate of HNSCC patients remains 40–50% due to tumor metastasis, recurrence, and drug resistance  $[4]$  $[4]$ . Therefore, searching for specific molecular markers of HNSCC and strengthing research into the molecular mechanism underlying its initiation and development are of paramount significance to further improve the early diagnosis and treatment of HNSCC [\[5–](#page-11-4)[7\]](#page-11-5).

Retinol-binding protein type 1 (RBP1) is a lipocalin protein family member that transports retinol from the liver to epithelial cells and provides the retina with retinol by specifically binding with the retinal epithelial cells [[8,](#page-11-6) [9](#page-11-7)]. Retinol is also known as vitamin A with implications for the proliferation and differentiation of epithelial cells [[10\]](#page-11-8). RBP1 controls conversion of retinol to retinyl esters and reduces the activity of retinyl esters, thereby affecting retinoic acid metabolism. During embryonic development, retinoic acid can inhibit carcinogenesis and regulate the growth and apoptosis of normal and aberrantly differentiated cells, demonstrating the potential involvement of retinoic acid in the initiation and development of cancer. The up-regulation of RBP1 could promote the proliferation of lung adenocarcinoma cell line A549, and contribute to epithelial-mesenchymal transition by up-regulate CK5, CK6, CK14, CK17, RAR-α and down regulate RARβ [[11](#page-11-9)]. It was reported that downregulated RBP1 was associated with the occurrence of prostate cancer, endometrial cancer, and ovarian cancer, etc., while upregulated RBP1 was correlated with the occurrence of lung adenocarcinoma and laryngeal cancer, etc  $[12–15]$  $[12–15]$  $[12–15]$  $[12–15]$ .

Wu et al. experimented with RBP1-deficient mice and found an increased death rate than normal mice, a trend toward reduced blood count and platelet, and development of myelofibrosis and spleen and liver enlargement in partial mice, [[16](#page-11-12)]. Gao et al. proved that RBP1 overexpression was relevant to the malignant phenotype of oral squamous cell carcinoma, which was attributed to the deactivation of the RBP1-CKAP4 axis-mediated autophagy [[9\]](#page-11-7). However, there are no relevant studies concerning the relation between RBP1 and HNSCC.

In this study, the expression of RBP1 in HNSCC was analyzed based on the RNA-seq data from TCGA database, and it was verified in clinical tissue samples with the PCR method. The correlation between RBP1 expression and the prognosis of HNSCC patients was analyzed via Kaplan-Meier survival analysis, univariate and multivariate analyses. In addition, a nomogram was plotted to discuss the potential diagnostic and prognostic values of RBP1. Gene set enrichment analysis (GSEA) was applied to explore the potential biological function of RBP1. Overall, this study identified that RBP1 is an independent prognostic factor of HNSCC.

# **Methods**

# **Data acquisition**

All original data were acquired from The Cancer Genome Atlas (TCGA) (<https://cancergenome.nih.gov/>) and Gene Expression Omnibus (GEO) ([https://www.ncbi.nlm.](https://www.ncbi.nlm.nih.gov/geo/) [nih.gov/geo/](https://www.ncbi.nlm.nih.gov/geo/)) databases. Level-3 HTSeq-FPKM data of HNSCC patients, including 44 normal and 502 tumor cases, were downloaded from TCGA data portal [\(http://](http://tcga-data.nci.nih.gov/tcga/) [tcga-data.nci.nih.gov/tcga/](http://tcga-data.nci.nih.gov/tcga/)), then were transformed into transcripts per million reads (TPM) using the equation TPM= [FPKM(i)/sum (FPKM all transcripts)]  $\times$  10<sup>6</sup> and log2-transformed for subsequent analyses [[17](#page-11-13)]. Gene expression data were divided into high and low groups according to the median expression levels of RBP1. Both TCGA and GEO were a public open database, the relevant information obtained from there did not require additional ethics approval.

# **Quantitative real-time PCR analysis**

Oral squamous cell carcinoma tissues and adjacent control tissues were collected from patients who underwent surgical procedures at Guiyang Stomatology Hospital from 2021 to 2022. Total RNA was extracted from using TRIzol Universal Reagent (TIANGEN, Beijing, China) according to the manufacturer's instructions. RNA concentrations were determined on a Nanodrop2000 Spectrophotometer (Thermo Fischer Scientific, Waltham, MA, USA). Total RNA (2.5 µg) was subjected to cDNA synthesis using a qScript cDNA SuperMix (Quanta Biosciences, Beverly, MA, USA) through the following consequent cycles: firstly at 25 ℃ for 5 min, followed by 42 ℃ for 30 min and finally at 85 ℃ for 5 min. A real-time PCR was performed to determine the mRNA levels of RBP1 and GAPDH using SYBR Green Master MIX (ABI, Vernon, CA, USA). Real-time PCR results were calculated using the  $2^{-\Delta\Delta cq}$  method [\[18\]](#page-11-14).

# **Enrichment analysis**

Expression profiles (HTSeq-Counts) were compared between the high RBP1 expression group and the low RBP1 expression group to identify DEG using Wilcoxon rank-sum test [[19\]](#page-11-15) in the R language-related software, DESeq2 (version 1.26.0). Differences with a |log2 fold change|>1 and adjusted P-value<0.05 were considered threshold values for identifying DEGs [[20](#page-11-16)]. Gene ontology (GO) enrichment and Kyoto encyclopedia of genes

and genomes (KEGG) pathway analyses of the 280 DEGs were performed by "ClusterProfiler" package [\[21](#page-11-17)] and visualized by the "ggplot2" package. In addition, the protein-protein interaction network of RBP1 co-expressed genes was visualized by STRING (<http://string-db.org>; version 11.5) with a minimum level of confidence>0.4 to analyze the functional interactions among proteins [\[22](#page-11-18)].

# **Immune infiltration analysis by single-sample GSEA (ssGSEA)**

Immune infiltration analysis of HNSCC samples was performed by the ssGSEA method using the GSVA package in R ([http://www.biocondutor.org/package/release/](http://www.biocondutor.org/package/release/bioc/html/GSVA.html) [bioc/html/GSVA.html](http://www.biocondutor.org/package/release/bioc/html/GSVA.html)) for 24 types of immune cells [[23\]](#page-11-19), including neutrophils, mast cells, eosinophils, macrophages, natural killer (NK) cells, CD56dim NK cells, CD56bright NK cells, dendritic cells (DCs), immature DCs (iDCs), activated DCs (aDCs), plasmacytoid DCs ( $pDCs$ ), T cells,  $CD8<sup>+</sup>$  T cells, T helper (Th) cells, Th1 cells, Th2 cells, Th17 cells, T follicular helper cells, regulatory T cells (Treg), central memory T cells (Tcm), effector memory T cells (Tem), gamma delta T cells (Tgd), cytotoxic cells, and B cells. The Estimation of tumor microenvironments using expression data (ESTIMATE, version 1.0.13), including estimatescore, immuneScore, stromalscore [\[24](#page-11-20)]. Spearman correlation was used to calculate the correlation amoung RBP1, immune checkpoints and mismatch [\[25](#page-11-21)]. Analyses were performed with  $R$  (3.6.0), and visualized with ggplot2 (3.3.3).

#### **Statistical analysis**

GraphPad Prism (GraphPad 9.0 Software) and R software (3.6.3) were used for data analysis. Wilcoxon rank-sum test and Wilcoxon rank signed test was used to analyze the expression of RBP1 in unpaired and paired samples, respectively [[26](#page-11-22)]. In addition, ROC analysis and the frequently-used method for binary assessment were conducted using the pROC package  $(1.17.0.1)$   $[27]$  $[27]$  to assess the diagnostic capability of RPB1 in head and neck cancer. The computed AUC value from 0.5 to 1 indicates the discriminative potential from 50–100% [[28\]](#page-11-24). Kaplan– Meier survival curve was used for survival analysis [[29\]](#page-11-25). Univariate and multivariate analyses were carried out based on Cox proportional hazard regression [\[30](#page-11-26)]. P-value<0.05 was considered statistically significant.

# **Results**

#### **Relative expression level of RBP1 in HNSCC**

RBP1 mRNA expression level was analyzed in 33 cancers based on TCGA database. RBP1 was significantly up-regulated in 12 cancers, including head and neck squamous cell carcinoma, esophageal and oropharyngeal (Fig. [1](#page-3-0)A-C). Furthermore, the RBP1 mRNA expression of HNSCC was also higher than normal tissues both from the GSE85319 and the results of qPCR (Fig. [1](#page-3-0)D, G). Unfortunately, GSE85319 is missing clinical information to perform validation regarding RBP1 in clinical parameters. In addition, RBP1 expression showed promising discriminative power to identify tumors from normal tissue, with AUC value of 0.887 for TCGA and 0.738 for validation group (GSE83519) (Fig. [1](#page-3-0)E, F).

#### **Association of RBP1 mRNA with clinical characteristics**

The relationship between RBP1 mRNA expression and clinical parameters in HNSCC was assessed by UAL-CAN [\[31\]](#page-12-0), which is a comprehensive and interactive web resource for analyzing TCGA transcriptome and clinical patient data by TPM format. The results showed that RBP1 were differentially expressed in patients of different cancer stages, nodal metastasis status, TP53 mutation status, tumor grade, HPV infection status (Fig. [2\)](#page-4-0). In addition, the high RBP1 expression was associated with T stage, histologic grade, age(*P*<0.05) (Table [1\)](#page-5-0).

#### **Prognostic value of RBP1 in HNSCC**

KM survival curves demonstrated that patients having higher-level RBP1 expression tend to have a poor prognosis performance in terms of progression-free survival (PFI), and disease-free survival (DSS), overall survival (OS) (Fig. [3A](#page-6-0)–C). To confirm the correlation of RBP1 expression with overall survival (OS), disease-specific survival (DSS) patients, the prognostic factors influencing were identified by Cox regression analysis. Multivariate analyses revealed that RBP1 was an independent prognostic factor for OS in patients with HNSCC (HR=1.962, 95% CI: 1.277–3.013, *P*=0.002). Furthermore, primary therapy outcome (HR=0.202, 95% CI: 0.123–0.332, *P*<0.001), lymphovascular invasion (HR=1.764, 95% CI: 1.111–2.801, *P*=0.016), radiation therapy (HR=0.529, 95% CI: 0.332–0.841, *P*=0.007), were also an independent prognostic factor for OS in patients (Table [2](#page-6-1)). Disease-specific survival (DSS) could better reflect the clinical benefit of disease-specific. Based on the multivariate analysis of prognostic factors, RBP1 was also an independent prognostic factor of DSS (HR=1.905, 95% CI: 1.143–3.174, *P*=0.013) (Table [3\)](#page-7-0). Moreover, within the nomogram, RBP1 expression was found to contribute a high data points (ranging from 0 to 100) compared with the other clinical variables, which was consistent with the results of multivariate Cox regression. The C-index of the nomogram was 0.755 with 1000 bootstrap replicates (95% confidence interval: 0.731–0.779). The bias-corrected line in the calibration plot was close to the ideal curve (i.e., the 45-degree line) [\[32](#page-12-1)], indicating good agreement between the predicted and observed values (Fig. [3](#page-6-0)D, E).

<span id="page-3-0"></span>

**Fig. 1** (**A**) RBP1 expression significantly up-regulated in multiple cancers including HNSCC. ACC, Adrenocortical carcinoma; BLCA, Bladder Urothelial Carcinoma; BRCA, Breast invasive carcinoma; CESC, Cervical squamous cell carcinoma and endocervical adeno carcinoma; CHOL, Cholangio carcinoma; COAD, Colon adenocarcinoma; DLBC, Lymphoid Neoplasm Diffuse Large B-cell Lymphoma; ESCA, Esophageal carcinoma; GBM, Glioblastoma multiforme; HNSC, Head and Neck squamous cell carcinoma; KICH, Kidney Chromophobe; KIRC, Kidney renal clear cell carcinoma; KIRP, Kidney renal papillary cell carcinoma; LAML, Acute Myeloid Leukemia; LGG, Brain Lower Grade Glioma; LIHC, Liver hepatocellular carcinoma; LUAD, Lung adenocarcinoma; LUSC, Lung squamous cell carcinoma; MESO, Mesothelioma; OV, Ovarian serous cystadenocarcinoma; PAAD, Pancreatic adenocarcinoma; PCPG, Pheochromocytoma and Paraganglioma; PRAD, Prostate adenocarcinoma; READ, Rectum adenocarcinoma; SARC, Sarcoma; SKCM, Skin Cutaneous Melanoma; STAD, Stomach adenocarcinoma; TGCT, Testicular Germ Cell Tumors; THCA, Thyroid carcinoma; THYM, Thymoma; UCEC, Uterine Corpus Endometrial Carcinoma; UCS, Uterine Carcinosarcoma; UVM, Uveal Melanoma. (**B**) RBP1 expression levels in HNSCC and matched normal tissues. (**C**) RBP1 expression levels in HNSCC and normal tissues. (**D**) RBP1 expression levels were significantly higher in the HNSCC tissues compared to the adjacent peritumoral tissues in GSE83519. (**E**)ROC analysis of RBP1 shows promising discrimination power between tumor and normal tissues based on TCGA. (**F**)ROC analysis of RBP1 shows promising discrimination power between tumor and normal tissues based on GSE83519. (**G**) the level of RBP1 mRNA expression in 6 different head and neck squamous cell carcinoma and normal normal tissues

#### **Gene set enrichment analysis**

Based on significant differences (P-value<0.05,  $FDR < 0.25$ ), GSEA was used to identify signaling pathways associated with HNSCC between the high and low RBP1 expression groups, including 265 positive

regulation pathways and 212 negative regulation pathways. The most significantly enriched pathways were the mitochondrial translation, peptide hormone biosynthesis, translation, CD22 mediated BCR regulation, creation

<span id="page-4-0"></span>

**Fig. 2** Association of RBP1 expression and clinical characteristics of HNSCC. (**A**) Expression of RBP1 in HNSCC based on individual cancer stages. (**B**) Expression of RBP1 in HNSCC based on nodal metastasis status. (**C**)Expression of RBP1 in HNSC based on TP53 mutation status. (**D**) Expression of RBP1 in HNSCC based on tumor grade. (E) Expression of RBP1 in HNSC based on HPV infection status

of C4 and C2 activators, FCGR activation in positive and negative respectively (Fig. [4\)](#page-7-1).

# **Correlation between RBP1 and immune infiltration, immune checkpoints, mismatch repair genes in HNSCC**

The association between RBP1 expression and immune cell infiltration was analyzed using single-sample Gene Set Enrichment Analysis (ssGSEA). RBP1 expression was negatively correlated with the abundance of aDC, B cells, CD8<sup>+</sup> T cells, Cytotoxic cells, NK CD56dim cells, pDC, T cells, T helper cells, Tcm, Tem, TFH, TReg, and was positively correlated with the abundance of Tgd. (Fig. [5\)](#page-8-0). Pearson's method was used to estimate the correlations between the RBP1 expression and immune checkpoint molecules, mismatch repair genes, tumor microenvironment. RBP1 was related to ImmuneScore,

<span id="page-5-0"></span>



ESTIMATEScore, StromalScore (*P*<0.05, Fig. [6](#page-8-1)C). Coexpression analysis of RBP1 and immune Checkpoint molecules indicated that RBP1 was significantly associated with PDCD1, CTLA4 etc. (*P*<0.05, Fig. [6](#page-8-1)A). Coexpression analysis of RBP1 and mismatch repair genes indicated that RBP1 was related to MLH1, MSH2, MSH6, PMS2 in HNSCC (*P*<0.05, Fig. [6](#page-8-1)B).

#### **Functional annotation and predicted signaling pathways**

The top ten co-expression genes of RBP1 were shown in Fig. [7A](#page-9-0), and the most related gene was LRAT [\[33](#page-12-2)], is considered to have a cancer-promoting effect. To better understand the functional implication of RBP1 in HNSCC from the 280 DEGs identified between the high and low expression groups, GO enrichment analysis was performed using the ClusterProfile package. Furthermore, 12 enriched terms were identified in the GO "biological process" category, the top four items were ammonium binding, neuropeptide receptor binding structural constituent of eye lens, peptide hormone receptor, binding (Fig. [7](#page-9-0)B); 14 enriched terms were identified in the GO "biological process" category, including visual perception, learning, regulation of sensory perception regulation of sensory perception of pain (Fig. [7](#page-9-0)C). 12 enriched terms within the "cellular component" category were associated with the activation of multiple proteins (Fig. [7D](#page-9-0)). These were analyzed for correlation of RBP1 and TP53, CDKN2A, CCND1, PTEN in TCGA database, that revealed RBP1 was positively correlated with CCND1 and negatively correlated with TP53, CDKN2A (Supplement Fig. 1).

# **Bioinformatical analysis**

The correlation between RBP1 expression and its mutation in pan-cancer was analyzed using cBioPortal, as revealed in Fig. [8A](#page-10-0). In 523 HNSCC cases, the genetic alteration was found in 26 cases, and the mutation rate was 4.97%. Besides, RBP1 alteration in HNSCC was associated with a shorter overall survival, implying that the genetic mutation of RBP1 could also affect HNSCC patients' prognosis (Fig. [8B](#page-10-0)). The methylation level of RBP1 in HNSCC was examined by UALCAN based on TCGA. The methylation was higher in normal group (Fig. [8C](#page-10-0)). MethSurv analysis showed that patients with high RBP1 methylation had a better overall survival than patients with low RBP1 methylation. The 6 CpG sites are shown in Fig. [8D](#page-10-0), each hazard ratio and 95% confidence interval are shown in supplement Table 1.

#### **Discussion**

Pan-cancer analysis demonstrated that RBP1 mRNA expression was significantly up-regulated in ACC, BRCA, CESC, DLBC, ESCA, GBM, HNSC, LUSC, THCA, THYM, UCEC, and UCS. PCR result showed significantly higher RBP1 mRNA expression in HNSCC tissues than adjacent normal tissues, and correlation analysis revealed a close relationship between high RBP1 expression and advanced cancer stage and tumor grade. TP53 plays a key regulatory role in apoptosis and cell cycle, and mutations in the TP53 gene are considered as the earliest and indispensable event in the initiation and development of HNSCC [34-[36](#page-12-4)]. TCGA data found that RBP1 expression was relevant to the TP53 mutation status in HNSCC patients. TP53, CDKN2A, CCND1, PTEN are considered important driver gene of head and neck squamous cell carcinoma [[37](#page-12-5), [38\]](#page-12-6). The above evidence indicates that

<span id="page-6-0"></span>

**Fig. 3** The association of RBP1 expression and prognosis in HNSCC. Kaplan-Meier plotter database analysis shows the differences in (**A**) Progress free interval; (**B**) Disease specific survival; (**C**) Overall survival; (**D**) nomogram integrates RBP1 and other prognostic factors in HNSCC from TCGA data; (**E**) The calibration plot of the nomogram

<span id="page-6-1"></span>



RBP1 is negatively correlated with tumor suppressors TP53, CDKN2A and positively correlated with cancerpromoting gene CCND1. This type of correlation imply close coordination between RBP1 and other oncogenes in the development of squamous cell carcinoma. Beyond this, the result indicates that high-risk HPV infection is one of the contributing factors for OSCCs. RBP1 had a much higher expression level in HPV negative (−) patients, this means is not typically associated with HPV infection, more experiments are needed to verify. Previous studies revealed that RBP1 affected cell differentiation and tumor progression through interfering with retinoic acid metabolism by decreasing retinol transport, preventing retinyl esters formation, and reducing retinoic acid receptors activity [[9,](#page-11-7) [39–](#page-12-7)[41\]](#page-12-8). It was also reported that high expression of RBP1 is associated with bladder cancer, tongue cancer and laryngeal squamous-cell carcinoma. Gao et al. proved that RBP1 overexpression was

<span id="page-7-0"></span>Table 3 Association of clinicopathological characteristics with disease-specific survival using univariate or multivariate Cox regression analysis



<span id="page-7-1"></span>

**Fig. 4** Ecrichment plots of RBP1 in HNSCC from GSEA. (**A**) Mitochondrial translation; (**B**) Peptide hormone biosynthesis; (**C**)Translation; (**D**) CD22 mediated BCR regulation; (**E**) Creation of C4 and C2 activator; (**F**) FCGR activation

relevant to the malignant phenotype of oral squamous cell carcinoma, which was attributed to the deactivation of the RBP1-CKAP4 axis-mediated autophagy [[9\]](#page-11-7). Taken together, high expression of RBP1 may be associated with the initiation and development of HNSCC.

The present study also identified the remarkable prognostic value of RBP1 for the survival outcomes of HNSCC patients. Kaplan-Meier analysis revealed that higher expression of RBP1 indicated worse OS, PFI, and DSS in HNSCC patients, implying RBP1's potential as a tumor gene. Moreover, RBP1 was confirmed as an independent prognostic factor for the OS and DSS of HNSCC patients via univariate and multivariate COX regression analyses [\[42](#page-12-9)]. In all, RBP1 might be a potential therapeutic target of HNSCC. Nomogram is generated from the multi-variable model that predicts the probability of the occurrence or outcome of a certain disease, which is reliable and effective [[43,](#page-12-10) [44\]](#page-12-11). It has been extensively applied in clinical prognosis and decision-making. In this study, a nomogram combining RBP1 expression level, primary therapy outcome, lymphovascular invasion, and gender was established, in which RBP1

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Fig. 5 Correlations between the relative abundance of 24 immune cells and RBP1 expression levels. The size of the dots represents the absolute Spearman's correlation coefficient values

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**Fig. 6** Associations between RBP1 expression and tumor immune infiltration. (**A**) Associations between RBP1 expression and immune checkpoint molecules in HNSCC from TCGA dataset; (**B**) Associations between RBP1 expression and mismatch repair genes in HNSCC from TCGA dataset; (**C**) Associations between RBP1 expression and tumor microenvironment in HNSCC from TCGA dataset; \**P*<0.05; \*\**P*<0.05; \*\*\**P*<0.001

expression level contributed most, indicating that RBP1 is an effective prognostic factor.

prediction model for HNSCC, and the C-index was 0.687 in their nomogram [[5\]](#page-11-4).

Generally, the the closer the value of the C-index is to 1, the higher the accuracy of the predictive ability of the nomogram. The C-index for OS prediction was 0.755 with higher credibility than our previous study about DCBLD1(C-index was 0.720) [\[7](#page-11-5)]. Liang constructed a

In addition, the present study also explored the role of mutation and promoter methylation of the RBP1 gene in HNSCC prognosis. RBP1 mutations ars associated with poor prognosis in HNSCC. DNA methylation is the covalent modification of cytosine at 5′ site catalysed by

<span id="page-9-0"></span>

**Fig. 7** Protein–protein interaction network and GO enrichment in HNSCC. (**A**) RBP1 interaction protein plot and description. (**B**) The top 5 GO enrichment terms in MF. (**C**) The top 5 GO enrichment terms in BP.(**D**) The top 5 GO enrichment terms in CC

DNA methyl transferases, which play an important role in regulating gene activity and transcript levels without changing gene sequence [[45\]](#page-12-12). Studies have reported that DNA methylation is associated with tumor occurrence and development [\[46,](#page-12-13) [47](#page-12-14)]. Chou observed that glioma patients with RBP1 hypermethylation are associated with a better prognosis, and the expression of RBP1 was associated with hypermethylation [[48\]](#page-12-15). We analyzed the methylation level of RBP1 in HNSCC and found that RPB1 promoter methylation was much more prevalent in normal tissue than in HNSCC tissue. Moreover, RBP1 methylation is associated with good prognosis of HNSCC, which was consisted with chou's study. Esteller reported that transcriptional silencing of RBP1 was associated with CpG island promoter hypermethylation in HN12(head and neck cancer cell lian), Shigeru Tsunoda et al. found that RBP1 showed more frequent methylation in the tumor than the matched proximal resection margin of uninvolved esophagus [[12](#page-11-10), [49\]](#page-12-16).

Tumor microenvironment is of vital importance in the development and treatment of tumors [[50\]](#page-12-17). Research showed that massive infiltration of immune cells in the tumor tissues was substantially associated with the prognosis of patients [[51\]](#page-12-18). The present study analyzed the relationships between RBP1 and the tumor microenvironment, tumor immune infiltration, immune checkpoint molecules, and mismatch repair genes. Previous studies found that the tumor infiltration of B cells and CD8<sup>+</sup> T cells was relevant to the poor prognosis of HNSCC patients [[52–](#page-12-19)[54\]](#page-12-20). Moreover, higher expression

of RBP1 led to shorter OS in patients, and RBP1 might impact the prognosis of HNSCC patients via affecting the tumor microenvironment.

However, the prediction of single gene still had some limitations and is obviously not as good as multigene. The initiation and progression of tumors result from the mutational synergism of genes with similar functions. In contrast, there was significant tumor suppression in monogenic therapy. For example, p53 was predictive of a cisplatin-based therapeutic benefit in patients with head and neck squamous cell carcinoma [[55](#page-12-21)]. Pembrolizumab as PD-L1 inhibitor have established clinical use. Therefore, single gene analysis still plays a significant role in anticancer treatment [[56](#page-12-22)].

To conclude, this study validated the significantly upregulated expression of RBP1 in HNSCC and identified it as an independent risk factor for the prognosis of HNSCC patients. RBP1, therefore, has potential research value in HNSCC targeted therapy.

#### **Conclusion**

The relationship between RBP1 expression and HNSCC was described for the first time based on TCGA database. Our study found that RBP1 is a potential prognostic factor for survival outcomes. Besides, the RBP1 methylation is correlated with better prognosis of HNSCC. RBP1 is expected to be a novel diagnostic and prognostic factor for HNSCC.

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**Fig. 8** Mutation feature of RBP1 in HNSCC from TCGA cohort using the cBioPortal tool; (**A**) The alteration frequency with mutation type of RBP1 in different tumor samples from TCGA cohorts; (**B**) K-M survival analysis of OS with or without RBP1 alteration; (**C**) The methylation level of RBP1 in HNSCC; (**D**) The Kaplan-Meier survival of the promoter methylation of RBP1S



# **Supplementary Information**

The online version contains supplementary material available at [https://doi.](https://doi.org/10.1186/s12885-024-12565-3) [org/10.1186/s12885-024-12565-3](https://doi.org/10.1186/s12885-024-12565-3).

Supplementary Material 1

Supplementary Material 2

## **Acknowledgements**

Not applicable.

#### **Author contributions**

LLF: conceived the study, supervised the experiments and drafted the manuscript. MY: conceived the study, supervised the experiments, and drafted the manuscript. XY, MS: data evaluation and manuscript preparation. MG, RE. F, TV, RS: analyzed the data and revised the manuscript. LYX: conceived the study, designed the data evaluation, and prepared the manuscript. HCF: conceived the study, designed the data evaluation, and prepared the manuscript.

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#### **Data availability**

The datasets analyzed during the current study are publicly available from the following online databases: TCGA (<https://tcga-data.nci.nih.gov/tcga/>).

#### **Declarations**

**Ethical approval and consent to participate** Not applicable.

#### **Consent for publication**

Not applicable in the declarations section.

#### **Competing interests**

The authors declare no competing interests.

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## **References**

- <span id="page-11-0"></span>1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68(6):394–424.
- <span id="page-11-1"></span>2. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer. 2015;136(5):E359–386.
- <span id="page-11-2"></span>3. Fortpied C, Vinches M. The statistical evaluation of treatment and outcomes in Head and Neck squamous cell carcinoma clinical trials. Front Oncol. 2019;9:634.
- <span id="page-11-3"></span>4. Ferlay J, Colombet M, Soerjomataram I, Mathers C, Parkin DM, Pineros M, Znaor A, Bray F. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. Int J Cancer. 2019;144(8):1941–53.
- <span id="page-11-4"></span>Liang L, Huang Q, Gan M, Jiang L, Yan H, Lin Z, Zhu H, Wang R, Hu K. High SEC61G expression predicts poor prognosis in patients with Head and Neck squamous cell carcinomas. J Cancer. 2021;12(13):3887–99.
- 6. Oliva M, Spreafico A, Taberna M, Alemany L, Coburn B, Mesia R, Siu LL. Immune biomarkers of response to immune-checkpoint inhibitors in head and neck squamous cell carcinoma. Ann Oncol. 2019;30(1):57–67.
- <span id="page-11-5"></span>7. Fu LL, Yan M, Ma MX, Luo Y, Shao M, Gosau M, Friedrich RE, Vollkommer T, Feng HC, Smeets R. DCBLD1 overexpression is Associated with a poor prognosis in Head and Neck squamous cell carcinoma. Front Immunol. 2022;13:939344.
- <span id="page-11-6"></span>8. Yang Q, Graham TE, Mody N, Preitner F, Peroni OD, Zabolotny JM, Kotani K, Quadro L, Kahn BB. Serum retinol binding protein 4 contributes to insulin resistance in obesity and type 2 diabetes. Nature. 2005;436(7049):356–62.
- <span id="page-11-7"></span>9. Gao L, Wang Q, Ren W, Zheng J, Li S, Dou Z, Kong X, Liang X, Zhi K. The RBP1- CKAP4 axis activates oncogenic autophagy and promotes cancer progression in oral squamous cell carcinoma. Cell Death Dis. 2020;11(6):488.
- <span id="page-11-8"></span>10. Thomas S, Prabhu R, Balasubramanian KA. Retinoid metabolism in the rat small intestine. Br J Nutr. 2005;93(1):59–63.
- <span id="page-11-9"></span>11. Doldo E, Costanza G, Ferlosio A, Pompeo E, Agostinelli S, Bellezza G, Mazzaglia D, Giunta A, Sidoni A, Orlandi A. High expression of cellular retinol binding protein-1 in lung adenocarcinoma is associated with poor prognosis. Genes Cancer. 2015;6(11–12):490–502.
- <span id="page-11-10"></span>12. Esteller M, Guo M, Moreno V, Peinado MA, Capella G, Galm O, Baylin SB, Herman JG. Hypermethylation-associated inactivation of the Cellular Retinolbinding-protein 1 gene in Human Cancer. Cancer Res. 2002;62(20):5902–5.
- 13. Jeronimo C, Henrique R, Oliveira J, Lobo F, Pais I, Teixeira MR, Lopes C. Aberrant cellular retinol binding protein 1 (CRBP1) gene expression and promoter methylation in prostate cancer. J Clin Pathol. 2004;57(8):872–6.
- 14. Toki K, Enokida H, Kawakami K, Chiyomaru T, Tatarano S, Yoshino H, Uchida Y, Kawahara K, Nishiyama K, Seki N, et al. CpG hypermethylation of cellular retinol-binding protein 1 contributes to cell proliferation and migration in bladder cancer. Int J Oncol. 2010;37(6):1379–88.
- <span id="page-11-11"></span>15. Orlandi A, Ferlosio A, Ciucci A, Francesconi A, Lifschitz-Mercer B, Gabbiani G, Spagnoli LG, Czernobilsky B. Cellular retinol binding protein-1 expression in endometrial hyperplasia and carcinoma: diagnostic and possible therapeutic implications. Mod Pathol. 2006;19(6):797–803.
- <span id="page-11-12"></span>16. Wu MY, Eldin KW, Beaudet AL. Identification of chromatin remodeling genes Arid4a and Arid4b as leukemia suppressor genes. J Natl Cancer Inst. 2008;100(17):1247–59.
- <span id="page-11-13"></span>17. Fernandez R, Marcet-Houben M, Legeai F, Richard G, Robin S, Wucher V, Pegueroles C, Gabaldon T, Tagu D. Selection following gene duplication shapes recent genome evolution in the pea aphid Acyrthosiphon pisum. Mol Biol Evol. 2020;37(9):2601–15.
- <span id="page-11-14"></span>18. Rao X, Huang X, Zhou Z, Lin X. An improvement of the 2^(-delta delta CT) method for quantitative real-time polymerase chain reaction data analysis. Biostat Bioinforma Biomath. 2013;3(3):71–85.
- <span id="page-11-15"></span>19. Love MI, Huber W, Anders S. Moderated estimation of Fold change and dispersion for RNA-seq data with DESeq2. Genome Biol. 2014;15(12):550.
- <span id="page-11-16"></span>20. Ji Y, Yin Y, Zhang W. Integrated Bioinformatic Analysis Identifies Networks and Promising Biomarkers for Hepatitis B Virus-Related Hepatocellular Carcinoma. Int J Genomics 2020, 2020:2061024.
- <span id="page-11-17"></span>21. Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. OMICS. 2012;16(5):284–7.
- <span id="page-11-18"></span>22. Szklarczyk D, Gable AL, Nastou KC, Lyon D, Kirsch R, Pyysalo S, Doncheva NT, Legeay M, Fang T, Bork P, et al. The STRING database in 2021: customizable protein-protein networks, and functional characterization of user-uploaded gene/measurement sets. Nucleic Acids Res. 2021;49(D1):D605–12.
- <span id="page-11-19"></span>23. Ma B, Li H, Qiao J, Meng T, Yu R. Immune-related miRNA signature identifies prognosis and immune landscape in head and neck squamous cell carcinomas. Biosci Rep 2020, 40(11).
- <span id="page-11-20"></span>24. Yoshihara K, Shahmoradgoli M, Martinez E, Vegesna R, Kim H, Torres-Garcia W, Trevino V, Shen H, Laird PW, Levine DA, et al. Inferring tumour purity and stromal and immune cell admixture from expression data. Nat Commun. 2013;4:2612.
- <span id="page-11-21"></span>25. Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC, Angell H, Fredriksen T, Lafontaine L, Berger A, et al. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. Immunity. 2013;39(4):782–95.
- <span id="page-11-22"></span>26. Natarajan S, Lipsitz SR, Fitzmaurice GM, Sinha D, Ibrahim JG, Haas J, Gellad W. An extension of the wilcoxon Rank-Sum test for complex sample survey data. J R Stat Soc Ser C Appl Stat. 2012;61(4):653–64.
- <span id="page-11-23"></span>27. Robin X, Turck N, Hainard A, Tiberti N, Lisacek F, Sanchez JC, Muller M. pROC: an open-source package for R and S + to analyze and compare ROC curves. BMC Bioinformatics. 2011;12:77.
- <span id="page-11-24"></span>28. Harrell FE Jr., Califf RM, Pryor DB, Lee KL, Rosati RA. Evaluating the yield of medical tests. JAMA. 1982;247(18):2543–6.
- <span id="page-11-25"></span>29. Stel VS, Dekker FW, Tripepi G, Zoccali C, Jager KJ. Survival analysis I: the Kaplan-Meier method. Nephron Clin Pract. 2011;119(1):c83–88.
- <span id="page-11-26"></span>30. Zhang Z, Reinikainen J, Adeleke KA, Pieterse ME, Groothuis-Oudshoorn CGM. Time-varying covariates and coefficients in Cox regression models. Ann Transl Med. 2018;6(7):121.
- <span id="page-12-0"></span>31. Chandrashekar DS, Karthikeyan SK, Korla PK, Patel H, Shovon AR, Athar M, Netto GJ, Qin ZS, Kumar S, Manne U, et al. UALCAN: an update to the integrated cancer data analysis platform. Neoplasia. 2022;25:18–27.
- <span id="page-12-1"></span>32. Song CK, Guo ZX, Shen XY, Wang YJ, Wang QW, Yu DH, Chen C, Liu XP, Huang JY, Li S et al. Prognostic Factors Analysis and Nomogram Construction of Dual Primary Lung Cancer: A Population Study. Biomed Res Int : 2020, 2020:7206591.
- <span id="page-12-2"></span>33. Cui GX, Gao XL, Liang XH. [Invasion and metastasis mechanism of human papillomavirus in head and neck squamous cell carcinomas]. Hua Xi Kou Qiang Yi Xue Za Zhi. 2018;36(5):544–51.
- <span id="page-12-3"></span>34. Gross AM, Orosco RK, Shen JP, Egloff AM, Carter H, Hofree M, Choueiri M, Coffey CS, Lippman SM, Hayes DN, et al. Multi-tiered genomic analysis of head and neck cancer ties TP53 mutation to 3p loss. Nat Genet. 2014;46(9):939–43.
- 35. Sun J, Yu X, Xue L, Li S, Li J, Tong D, Du Y. TP53-Associated Ion Channel genes serve as prognostic predictor and therapeutic targets in Head and Neck squamous cell carcinoma. Technol Cancer Res Treat. 2020;19:1533033820972344.
- <span id="page-12-4"></span>36. Chari NS, Ivan C, Le X, Li J, Mijiti A, Patel AA, Osman AA, Peterson CB, Williams MD, Pickering CR, et al. Disruption of TP63-miR-27a\* Feedback Loop by Mutant TP53 in Head and Neck Cancer. J Natl Cancer Inst. 2020;112(3):266–77.
- <span id="page-12-5"></span>37. Brown NA, Plouffe KR, Yilmaz O, Weindorf SC, Betz BL, Carey TE, Seethala RR, McHugh JB, Tomlins SA, Udager AM. TP53 mutations and CDKN2A mutations/deletions are highly recurrent molecular alterations in the malignant progression of sinonasal papillomas. Mod Pathol. 2021;34(6):1133–42.
- <span id="page-12-6"></span>Parikh AS, Yu VX, Flashner S, Okolo OB, Lu C, Henick BS, Momen-Heravi F, Puram SV, Teknos T, Pan Q, et al. Patient-derived three-dimensional culture techniques model tumor heterogeneity in head and neck cancer. Oral Oncol. 2023;138:106330.
- <span id="page-12-7"></span>39. Monroe DG, Hawse JR, Subramaniam M, Spelsberg TC. Retinoblastoma binding protein-1 (RBP1) is a Runx2 coactivator and promotes osteoblastic differentiation. BMC Musculoskelet Disord. 2010;11:104.
- 40. Binda O, Roy JS, Branton PE. RBP1 family proteins exhibit SUMOylationdependent transcriptional repression and induce cell growth inhibition reminiscent of senescence. Mol Cell Biol. 2006;26(5):1917–31.
- <span id="page-12-8"></span>41. Urvalek A, Laursen KB, Gudas LJ. The roles of retinoic acid and retinoic acid receptors in inducing epigenetic changes. Subcell Biochem. 2014;70:129–49.
- <span id="page-12-9"></span>42. Li M, Liang M, Lan T, Wu X, Xie W, Wang T, Chen Z, Shen S, Peng B. Four Immune-Related Long non-coding RNAs for prognosis prediction in patients with Hepatocellular Carcinoma. Front Mol Biosci. 2020;7:566491.
- <span id="page-12-10"></span>43. Nieder C, Mehta MP, Geinitz H, Grosu AL. Prognostic and predictive factors in patients with brain metastases from solid tumors: a review of published nomograms. Crit Rev Oncol Hematol. 2018;126:13–8.
- <span id="page-12-11"></span>44. Balachandran VP, Gonen M, Smith JJ, DeMatteo RP. Nomograms in oncology: more than meets the eye. Lancet Oncol. 2015;16(4):e173–180.
- <span id="page-12-12"></span>Skvortsova K, Stirzaker C, Taberlay P. The DNA methylation landscape in cancer. Essays Biochem. 2019;63(6):797–811.
- <span id="page-12-13"></span>46. Lin YL, Chen HL, Cheng SB, Yeh DC, Huang CC, P'Eng FK, Tsai TC, Wu CC, Chen CM. Methylation-silencing RCC1 expression is associated with tumorigenesis and depth of invasion in gastric cancer. Int J Clin Exp Pathol. 2015;8(11):14257–69.
- <span id="page-12-14"></span>47. Brumbaugh J, Kim IS, Ji F, Huebner AJ, Di Stefano B, Schwarz BA, Charlton J, Coffey A, Choi J, Walsh RM, et al. Inducible histone K-to-M mutations are dynamic tools to probe the physiological role of site-specific histone methylation in vitro and in vivo. Nat Cell Biol. 2019;21(11):1449–61.
- <span id="page-12-15"></span>48. Chou AP, Chowdhury R, Li S, Chen W, Kim AJ, Piccioni DE, Selfridge JM, Mody RR, Chang S, Lalezari S, et al. Identification of retinol binding protein 1 promoter hypermethylation in isocitrate dehydrogenase 1 and 2 mutant gliomas. J Natl Cancer Inst. 2012;104(19):1458–69.
- <span id="page-12-16"></span>49. Tsunoda S, Smith E, De Young NJ, Wang X, Tian ZQ, Liu JF, Jamieson GG, Drew PA. Methylation of CLDN6, FBN2, RBP1, RBP4, TFPI2, and TMEFF2 in esophageal squamous cell carcinoma. Oncol Rep. 2009;21(4):1067–73.
- <span id="page-12-17"></span>50. Binnewies M, Roberts EW, Kersten K, Chan V, Fearon DF, Merad M, Coussens LM, Gabrilovich DI, Ostrand-Rosenberg S, Hedrick CC, et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. Nat Med. 2018;24(5):541–50.
- <span id="page-12-18"></span>51. Bian T, Zheng M, Jiang D, Liu J, Sun H, Li X, Liu L, Zhang J, Liu Y. Prognostic biomarker TUBA1C is correlated to immune cell infiltration in the tumor microenvironment of lung adenocarcinoma. Cancer Cell Int. 2021;21(1):144.
- <span id="page-12-19"></span>52. So YK, Byeon SJ, Ku BM, Ko YH, Ahn MJ, Son YI, Chung MK. An increase of CD8(+) T cell infiltration following recurrence is a good prognosticator in HNSCC. Sci Rep. 2020;10(1):20059.
- 53. Zhang S, Wang B, Ma F, Tong F, Yan B, Liu T, Xie H, Song L, Yu S, Wei L. Characteristics of B lymphocyte infiltration in HPV(+) head and neck squamous cell carcinoma. Cancer Sci. 2021;112(4):1402–16.
- <span id="page-12-20"></span>54. Sandoval F, Terme M, Nizard M, Badoual C, Bureau MF, Freyburger L, Clement O, Marcheteau E, Gey A, Fraisse G, et al. Mucosal imprinting of vaccineinduced CD8(+) T cells is crucial to inhibit the growth of mucosal tumors. Sci Transl Med. 2013;5(172):172ra120.
- <span id="page-12-21"></span>55. Sobol RE, Menander KB, Chada S, Wiederhold D, Sellman B, Talbott M, Nemunaitis JJ. Analysis of Adenoviral p53 gene therapy clinical trials in recurrent Head and Neck squamous cell carcinoma. Front Oncol. 2021;11:645745.
- <span id="page-12-22"></span>56. Newton HS, Gawali VS, Chimote AA, Lehn MA, Palackdharry SM, Hinrichs BH, Jandarov R, Hildeman D, Janssen EM, Wise-Draper TM et al. PD1 blockade enhances K(+) channel activity, ca(2+) signaling, and migratory ability in cytotoxic T lymphocytes of patients with head and neck cancer. J Immunother Cancer 2020, 8(2).

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