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JMJD6 functions as an oncogene and is associated with poor prognosis in esophageal squamous cell carcinoma

Honggang Liu^{1†}, Menglong Jiang^{2†}, Fenghui Ma^{3†}, Jiapei Qin⁴, Xin Zhou⁴, Liquan Xu^{5*}, Xiaolong Yan^{1*} and Tao Jiang^{1*}

Abstract

Background Esophageal squamous cell carcinoma (ESCC) is one of the most common malignant tumors with a high prevalence and poor prognosis. It is an urgent problem to deeply understand the molecular mechanism of ESCC and develop effective diagnostic and prognostic methods.

Methods Using tumor tissue and corresponding paracancerous samples from 141 resected ESCC patients, we assessed Jumonji domain-containing protein 6 (JMJD6) expression using Immunohistochemical (IHC) staining. Kaplan-Meier survival analysis and univariate or multivariate analysis were used to investigate the relationship between JMJD6 expression and clinicopathological features. The expression status and prognostic value of JMJD6 were analyzed by bioinformatics and enrichment analysis.

Results The expression of JMJD6 in ESCC samples was higher than that in the corresponding paracancerous samples, and high expression of JMJD6 was positively associated with poor prognosis of ESCC patients. In addition, bioinformatics analysis of the expression and prognosis of JMJD6 in a variety of tumors showed that high expression of JMJD6 was significantly associated with poor overall survival (OS) in ESCC patients. Enrichment analysis indicated that the high expression of genes similar to JMJD6, such as *Conserved oligomeric Golgi 1 (COG1)*, *Major facilitator superfamily domain 11 (MFSD11)* and *Death Effector Domain Containing 2 (DEDD2)*, was associated with poor prognosis of ESCC, suggesting that JMJD6 might be involved in the occurrence and prognosis of ESCC.

Conclusion Our study found that JMJD6 expression was significantly increased in ESCC patients and positively correlated with prognosis, indicating that targeting JMJD6 might be an attractive prognostic biomarker and provides a potential treatment strategy for ESCC.

Trial registration The study was approved by Tangdu Hospital ethics committee (No. TDLL-202110-02).

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Keywords Esophageal squamous cell carcinoma, Jumonji domain-containing protein 6, Immunohistochemical staining, Prognosis

Background

Esophageal cancer, the sixth leading cause of cancer-related death worldwide, is a complex disease whose etiology varies by histologic type and population [1]. Esophageal squamous cell carcinoma (ESCC) is the main histological subtype with an extremely high prevalence and poor prognosis in Asia, accounting for about 90% of the incidence of esophageal cancer [2]. In recent years, with the improvement of diagnosis and treatment of esophageal cancer, the 5-year survival rate of ESCC patients has been greatly improved [3]. However, the molecular mechanism by which JMJD6 promotes the progression of ESCC remains unclear and warrants further exploration to develop promising therapeutic approaches.

Jumonji domain-containing protein 6 (JMJD6), also known as phosphatidylserine receptor, is a member of Jumonji C domain-containing proteins [4]. JMJD6 has arginine demethylase and lysyl hydroxylase activities in both histone and non-histone proteins [5, 6]. Furthermore, JMJD6 functions as a tyrosine kinase of histones, suggesting that JMJD6 acts at the transcriptional, splicing, posttranscriptional, and biochemical levels through chromatin configuration and epigenetic regulation [7]. Many studies have confirmed the role of JMJD6 in infection, inflammation, immunity, placental angiogenesis, tissue differentiation and embryonic development under normal physiological conditions [7, 8]. Abnormal expression of JMJD6 may contribute to the development of many diseases, such as neuropathic pain, foot-and-mouth disease, gestational diabetes, hepatitis C, and various types of cancer [7, 9–12]. Studies have shown that JMJD6, as an oncogene, is involved in the pathogenesis of many cancers, including oral cancer [13], colorectal cancer [14], breast cancer [6], lung cancer [15], and hepatocellular carcinoma [16] etc., and is closely related to the poor prognosis of patients. In addition, subsequent studies have demonstrated that JMJD6 has been implicated in the resistance of doxorubicin, methotrexate, etoposide and other chemotherapy drugs [7]. However, the role and molecular mechanism of JMJD6 in esophageal cancer, especially in ESCC, remain unclear. It would be important to continue efforts to elucidate the physiological functions of JMJD6 and the mechanisms by which JMJD6 contributes to ESCC progression.

To our knowledge, there has been no comprehensive analysis of the functional and clinical importance of JMJD6 in ESCC. In this study, we firstly systematically analyzed the expression status and prognostic value of JMJD6 by bioinformatics analysis in public datasets. Then

we assessed JMJD6 expression and prognosis by immunohistochemical (IHC) staining of 141 paired ESCC samples and clinicopathological characteristics. Finally, Enrichment analyses found that JMJD6 may be involved in the occurrence and prognosis of ESCC. Our findings demonstrated that the expression of JMJD6 was significantly increased in ESCC patients and positively correlated with prognosis in ESCC. Given the important role of JMJD6 in cancer progression, we suggest that investigating JMJD6 may become an attractive diagnostic and prognosis biomarker and potential strategy to develop novel cancer therapeutics.

Methods

Database-mining

Data from the Gene Expression Profiling Interactive Analysis version 2 (GEPIA2) database (<http://gepia2.cancer-pku.cn/>) was analyzed, including JMJD6 expression, survival analysis, and correlation analysis. Furthermore, we used the Kaplan-Meier plotter database (<https://kmpplot.com/>) to analyze the relationship between gene expression and prognosis of ESCC patients. Using STRING tool (version 11.5) (<https://string-db.org/>) to create a co-expression network of JMJD6.

Gene expression analysis of JMJD6 and survival prognosis analysis

GEPIA2 is for analyzing RNA-sequencing data of tumor and normal tissue samples from the TCGA and Genotype-Tissue Expression projects using a standard processing pipeline [17]. A survival significance map of JMJD6 in various types of cancer, as well as overall survival (OS) Kaplan–Meier plots, was generated using the GEPIA2 “Survival Analysis” module. The expression threshold was 75% for high JMJD6 expression and 25% for low JMJD6 expression.

The Kaplan-Meier plotter can assess the impact of 54,000 genes on survival in a variety of cancer types. Gene expression and OS were downloaded from the Gene Expression Omnibus, European Genome-phenome Archive, and TCGA databases [18]. The purpose of this tool is to provide benefits in clinical decisions, healthcare policy and resource allocation based on a meta-analysis of biomarker assessments. The correlation between gene expression and survival in ESCC was analyzed using the Kaplan-Meier plotter. Hazard ratios (HR) with 95% confidence intervals and Logrank *p* values were also calculated.

Patients and tissue samples

All ESCC cancer tissues and paired adjacent non-cancerous tissues were obtained from patients who underwent surgery of ESCC at Tangdu Hospital from January 2012 to June 2015. None of the patients received radiotherapy or chemotherapy before surgery, and the last follow-up was updated until death or June 2020. These ESCC cancer patients were diagnosed and graded on the basis of pathological features. The study was approved by the above-mentioned hospital ethics committee (No. TDLL-202110-02). This study was a retrospective study, and informed consent to participate was waived by Institution Review Board of Tangdu Hospital, Air Force Medical University.

IHC

The 141 pairs of ESCC tissues and corresponding tumor-adjacent normal tissues were placed into a paraffin-embedded tissue microarray. The primary antibody of anti-JMJD6 (1:1000, GB11341, Servicebio) was used for IHC staining. We applied the colon tissue section known to express JMJD6 as a positive control, and our IHC staining result was consistent with that of the Human Protein Atlas. As a negative control, we used sections of thymus tissue that were negative for JMJD6 expression. The positive and negative controls that have been run simultaneously with our study samples during IHC staining, which are shown in Fig. S3a. The IHC intensity scores included negative (score 0), weak (score 1), moderate (score 2), or strong staining (score 3). The proportion of positive stains was scored as 0 (<5%), 1 (6–25%), 2 (26–50%), 3 (51–75%), or 4 (>75%). The diagram of each staining pattern in IHC intensity scores (0/1/2/3) in tumor-adjacent normal/cancer tissue is shown in Fig. S3b. Multiply the two scores to get the total score. IHC staining was read by expert pathologists, and all pathologists were blinded to clinical data prior to statistical analysis. ESCC samples were divided into low or high JMJD6 expression groups according to their respective mean scores.

JMJD6-Related gene enrichment analysis

The STRING tool was used to create a co-expression network of Homo sapiens JMJD6. The main parameters were as follows: Active interaction source: network; meaning of network edges: evidence; maximum number of interactors: 100; and minimum required interaction score: high confidence (0.700). The platform generated graphs showing the top 14 proteins significantly co-expressed with JMJD6 expression in different types of cancer. The GEPIA2 “Similar Gene Detection” module was used to extract the 100 genes with the most similar expression patterns to JMJD6. Using the gene symbols of these 100 genes as input gene symbols in OmicShare, gene ontology pathway enrichment analysis was performed for

biological processes, cellular components, and molecular functional categories. In addition, gene correlation analysis was performed using the GEPIA2 “Correlation Analysis” module.

Statistical analysis

SPSS 23.0 software was used for statistical analysis. The relationship between JMJD6 expression and clinicopathological features in patients with ESCC was evaluated by the χ^2 test or Fisher’s exact test. Survival analyses were assessed using the Kaplan-Meier method and Log-rank test. Cox proportional hazards model was used for univariate and multivariate survival analysis. The student’s t-test was used for comparison between two groups. Data was presented as mean \pm standard deviation (SD). A value of $p < 0.05$ was considered as statistically significant difference.

Results

Correlation of JMJD6 expression and ESCC prognosis and survival

To assess the association between JMJD6 and prognosis, we investigated human tumors using GEPIA2. The relationship between JMJD6 and prognosis varied across different types of cancers. In most cancers, higher JMJD6 expression was associated with shorter OS in cases of adrenocortical carcinoma (ACC) (HR=5.3, Logrank $p=0.017$), esophageal carcinoma (ESCA) (HR=2.1, Logrank $p=0.026$), kidney renal clear cell carcinoma (KIRC) (HR=2.4, Logrank $p=0.00015$), kidney renal papillary cell carcinoma (KIRP) (HR=3.3, Logrank $p=0.014$), brain lower grade glioma (LGG) (HR=2.1, Logrank $p=0.0033$), liver hepatocellular carcinoma (LIHC) (HR=2.2, Logrank $p=0.00079$), mesothelioma (MESO) (HR=4.4, Logrank $p=5.9e-05$) (Fig. 1a and b and Fig. S1). In addition, based on datasets of GEPIA2, we found that JMJD6 was highly expressed in ESCA compared with non-tumor tissues ($p < 0.05$) (Fig. 1c). As ESCC is the main histological subtype of ESCA and the prevalence is extremely high in Asia, especially in China, we further used Kaplan-Meier plotter to analyze survival for confirmation of the association between JMJD6 and prognosis of ESCC. Patients in the JMJD6-high group had shorter OS than those in the JMJD6-low group with ESCC (HR=3.82 [1.09–13.36], Logrank $p=0.024$) (Fig. 1d). These findings indicated that JMJD6 expression was elevated in ESCC and it most likely served as a biomarker for poor prognosis.

To verify the role of JMJD6 in ESCC, we analyzed whether its expression was correlated with clinicopathological variables. We performed Cox proportional hazard regression analysis of the patients’ OS. Univariate analysis showed that JMJD6 expression (HR=2.282 [1.509–3.451], $p < 0.001$), lymphatic metastasis (HR=1.806

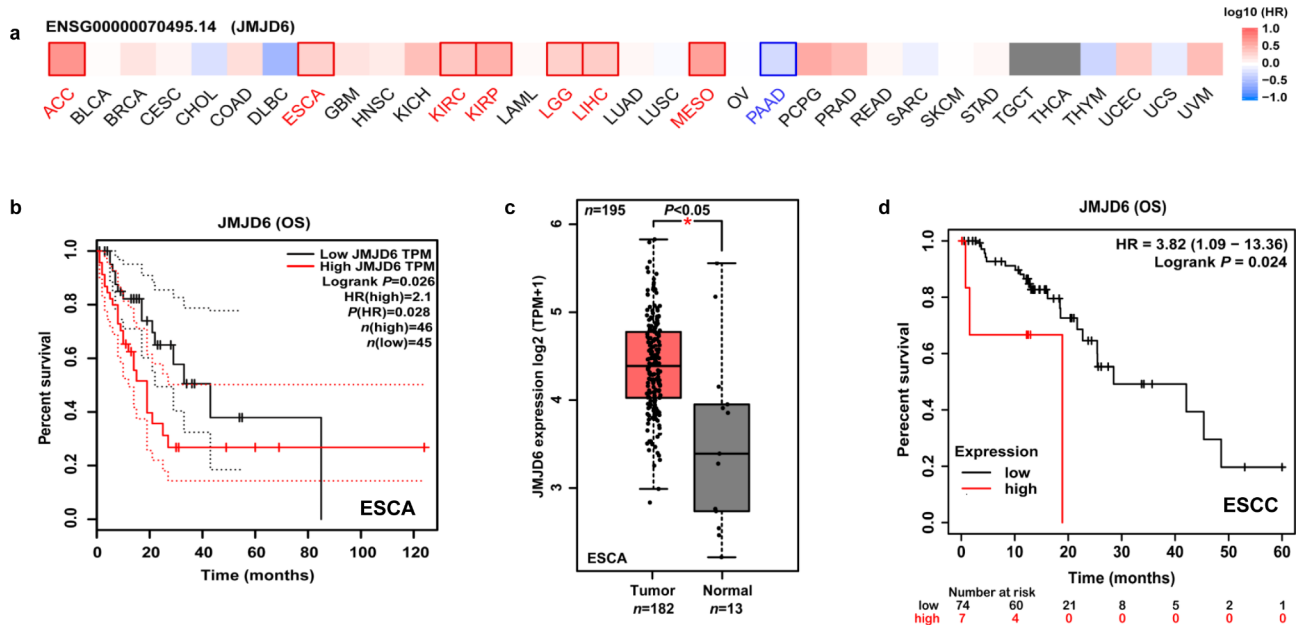


Fig. 1 Correlation between JMJD6 expression and OS in patients with different tumor types. GEPIA2 was used to construct (a) a survival map and perform (b) OS analysis, as well as (c) a statistical plot of JMJD6 expression in ESCA and non-tumor tissues. (d) Kaplan-Meier plotter analysis showed that ESCC patients in the JMJD6-high group had shorter OS than those in the JMJD6-low group. The survival map and Kaplan–Meier plots with significant results are shown. The 95% confidence intervals of overall survival are indicated by red and black dotted lines for high and low JMJD6 groups, respectively. Abbreviations: ESCA, esophageal carcinoma; ESCC, Esophageal squamous cell carcinoma; GEPIA2, Gene Expression Profiling Interactive Analysis version 2; JMJD6, Jumonji domain-containing protein 6; OS, overall survival

Table 1 Univariate and multivariate analyses of prognostic factors for ESCC patient survival

Variables	Univariate analysis			Multivariate analysis		
	HR	95% CI	P value	HR	95% CI	P value
Age (≥ 60 years/<60 years)	1.265	0.814–1.965	0.296			
Gender (Male/Female)	1.051	0.622–1.775	0.853			
Differentiation (Poor/Well and moderate)	1.027	0.560–1.884	0.930			
Lymphatic metastasis (N1-N3/N0)	1.806	1.204–2.710	0.004	1.412	0.507–3.934	0.509
pTNM stage (III/I-II)	1.731	1.150–2.605	0.008	1.140	0.407–3.189	0.803
JMJD6 expression (High/Low)	2.282	1.509–3.451	<0.001	2.113	1.388–3.218	<0.001

[1.204–2.710], $p=0.004$), and tumor-node-metastasis (TNM) stage (HR=1.731 [1.150–2.605], $p=0.008$) were associated with ESCC patients’ survival (Table 1). Furthermore, multivariate Cox regression analyses showed that JMJD6 was an independent prognostic factor for ESCC patients (HR=2.113 [1.388–3.218], $p<0.001$, Table 1). These results demonstrated that higher JMJD6 expression was associated with poor prognosis in patients with ESCC and the promotion of lymph node metastasis might be an important factor.

Expression of JMJD6 in ESCC patients

To further validate the JMJD6 protein level in ESCC tissues, we detected JMJD6 expression using IHC analysis in a tissue microarray containing 141 paired ESCC tumor-normal tissues (Fig. 2a). As shown in Fig. S3, sections of colon tissue that is known to express JMJD6 were used as a positive control, and sections of thymic tissue that is

negative for JMJD6 were used as a negative control, and each staining pattern in IHC intensity score (0/1/2/3) of tumor-adjacent normal/cancer tissue is shown. The expression of JMJD6 in ESCC tissues was significantly higher than that in adjacent noncancerous samples ($p<0.05$, Fig. 2b). In addition, ESCC patients with lymph node metastasis (N1-3), poorly differentiated, and a high American Joint Committee on Cancer (AJCC) 7th stage (stage III) had a significantly higher JMJD6 expression than those without lymph node metastasis (N0), with well and moderate differentiation and a lower 7th AJCC stage (stage I/II) ($p<0.05$, Table 2). Kaplan–Meier analysis also showed that ESCC patients with high JMJD6 expression had a shorter OS (HR=2.215 [1.470–3.336], Logrank $p<0.001$) (Fig. 2c). Furthermore, our results also indicated that the prognosis of ESCC patients with high JMJD6 expression was poor in the late clinical stage (stage III) (HR=2.306 [1.366–3.894], Logrank $p=0.002$).

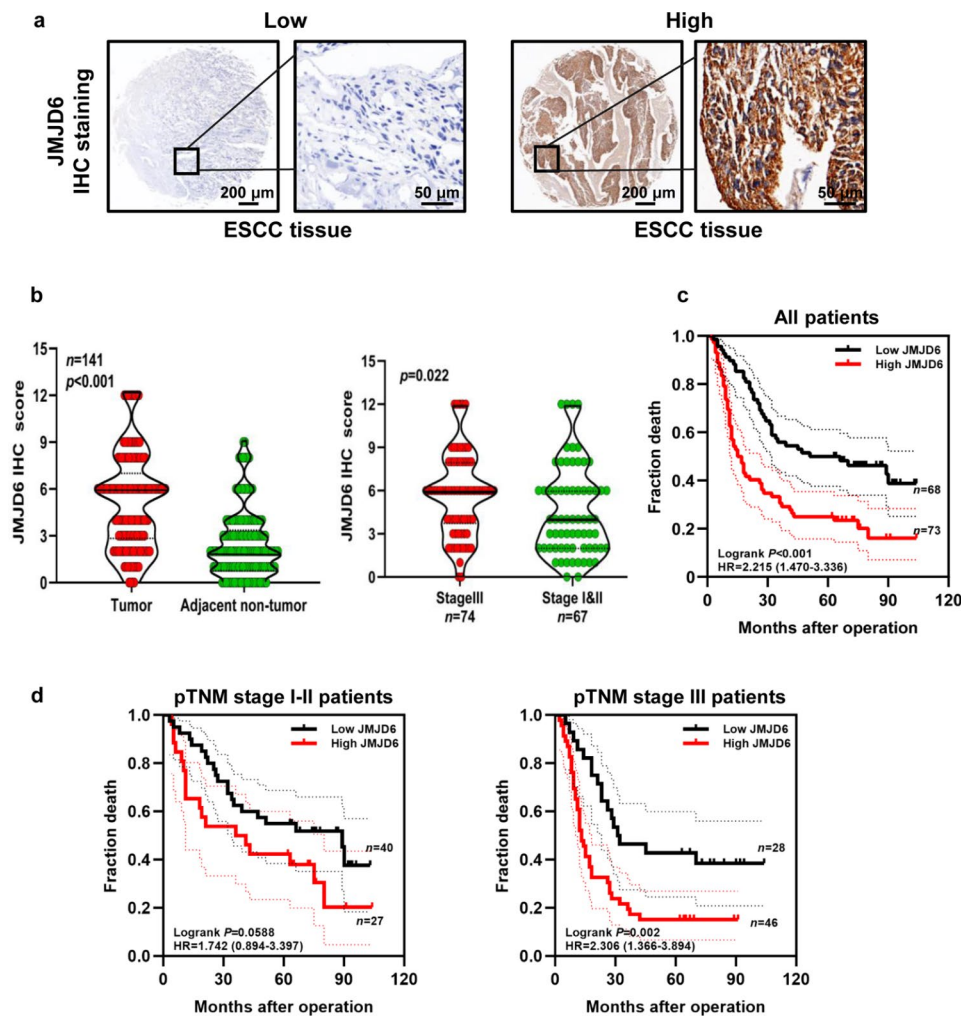


Fig. 2 High JMJD6 expression in ESCC tissues is associated with poor prognosis. **(a)** Representative IHC image of JMJD6 expression in ESCC. Scale bars, 200 and 50 μ m (inset). **(b)** The expression of JMJD6 in 141 patients with ESCC was statistically analyzed by IHC staining results based on tissue microarray. **(c-d)** Kaplan-Meier survival analyses of high or low JMJD6 expression based on tissue microarray IHC results for 141 patients **(c)**, 67 ESCC patients in the early clinical stages, and 74 ESCC patients in the late clinical stage **(d)**. Abbreviations: ESCC, Esophageal squamous cell carcinoma; IHC, Immunohistochemical; JMJD6, Jumonji domain-containing protein 6

However, there was no significant difference in prognosis in the early clinical stage (stage I/II) (HR=1.742 [0.894–3.397], Logrank $p=0.0588$) (Fig. 2d).

JMJD6-Related gene enrichment analysis

To investigate the functional mechanism of JMJD6 in carcinogenesis, we used STRING tool to extract the top 14 proteins co-expressed with JMJD6. JMJD6 physically interacts with bromodomain-containing protein 4 (BRD4), mediator of RNA polymerase II transcription subunit 12 homolog (MED12), cyclin-dependent kinase 9 (CDK9), lysine demethylase 8 (KDM8), TP53 and etc. (Fig. 3a), which have well-characterized functions in tumorigenesis [19–23]. For example, BRD4 plays an important role in super-enhancer organization and regulation of oncogene expression. Furthermore, MED12 and BRD4 could cooperate to sustain

cancer growth upon loss of mediator kinase [24]. Following that, we used GEPIA2 “Similar Gene Detection” module to extract the top 100 genes with expression patterns similar to JMJD6 from ESCA in the TCGA datasets. Gene Ontology enrichment analysis indicated that these genes were closely linked to protein binding and protein serine/threonine kinase activity (Fig. 3b). The top 20 genes with the most similar expression patterns to JMJD6 in ESCA were shown and the prognostic value of these genes was explored in the website of Kaplan-Meier plotter (Fig. 3c and Fig. S4). A high expression of *Conserved oligomeric Golgi 1 (COG1)* (HR=2.26 [1-5.12], Logrank $p=0.045$), *Major facilitator superfamily domain 11 (MFSD11)* (HR=3.47 [1.52–7.91], Logrank $p=0.0017$), *Death Effector Domain Containing 2 (DEDD2)* (HR=2.31 [1.03–5.16], Logrank $p=0.036$) while a low expression of *Small nucleolar RNA host gene 16 (SNHG16)* (HR=2.5

Table 2 Association of JMJD6 expression with clinicopathological parameters of patients with ESCC

Category	JMJD6 expression			pvalue
	n	low	high	
Age				0.777
<60	44	22	22	
>=60	97	46	51	
Gender				0.841
Male	115	55	60	
Female	26	13	13	
Tumor invasion				0.464
T1-T2	12	7	5	
T3-T4	129	61	68	
Lymphatic metastasis				0.005
N0	74	44	30	
N1-N3	67	24	43	
Differentiation				0.040
Well and moderate	122	63	59	
Poor	19	5	14	
pTNM stage				0.009
I-II	67	40	27	
III	74	28	46	

[1–5.9], Logrank $p=0.033$), *USP36* (HR=2.86 [1.3–6.5], Logrank $p=0.0087$) was related to a worse OS in ESCC patients (Fig. 3d). However, according to the “Correlation Analysis” module of GEPIA2, we found that the expression of JMJD6 was strongly positively correlated with the expression levels of *COG1*, *MFS11*, *DEDD2*, *SNHG16* and *USP36* in ESCA (Fig. 3e). Based on these results, we speculated that JMJD6 might play a tumor-promoting role in ESCC.

Discussion

Despite improvements in treatment efficacy over the last 10 years, the diagnostic efficiency and the prognosis of patients with ESCC are still poor due to limited biomarkers and tumor aggressiveness. In order to find new biomarkers for early diagnosis and prognosis, we comprehensively analyzed the clinical value and prognostic role of JMJD6 in a variety of cancers, especially in ESCC by mining the database. Our results demonstrated that the expression of JMJD6 was significantly increased compared with normal tissue, and was positively correlated with lymph node metastasis and TNM stage. In addition, we used bioinformatics analysis to find that ESCC patients with high JMJD6 expression had a shorter OS. However, both GEPIA2 database and Kaplan-Meier plotter database showed no correlation between JMJD6 expression and RFS in ESCC patients (Fig S2). We further reviewed the literature and found high JMJD6 expression was associated with improved OS and RFS in cholangiocarcinoma, which indicated that JMJD6 was one of the

improved independent prognostic factors of OS and RFS [25].

JMJD6 is an Fe(II)- and 2-oxoglutarate-dependent oxygenase and plays a crucial role in the differentiation of various tissues and cells during embryogenesis, and it knockout exhibited abnormal neurodevelopmental phenotypes in mice [16]. Although the exact mechanism by which JMJD6 promotes tumorigenesis and progression has not been elucidated, the interaction of JMJD6 with cancer-related signaling pathways has been identified as one of the potential mechanisms. Previously study have shown that JMJD6 expression was higher in human oral squamous cell carcinoma by inducing interleukin 4 transcription and binding to its promoter region [13]. Inhibition of JMJD6 could restore hepatocyte nuclear factor 4 alpha levels in protein arginine methyl transferase 1-knockout hepatocytes and prevent increased hepatocyte proliferation [26]. Furthermore, JMJD6 could form protein complexes with N-Myc and BRD4 and regulate the transcription of E2F2, N-Myc and c-Myc. Knockdown of JMJD6 inhibited the proliferation and survival of neuroblastoma cells and tumor progression in mice [27]. Furthermore, high-level expression of JMJD6 in human neuroblastoma and lung adenocarcinoma (EAC) independently predicted poor patient prognosis [27, 28]. Our findings indicated that the expression of JMJD6 was increased in ESCC and higher JMJD6 expression was associated with poor prognosis. Furthermore, ESCC patients with high JMJD6 expression have a poor prognosis in the late clinical stage, but there is no significant difference in the early clinical prognosis. A recent study found that the expression of JMJD6 in prostate cancer was up-regulated with the progression of stage and grade [29]. High expression of JMJD6 was closely related to advanced clinicopathologic stage, strong invasiveness and poor prognosis of melanoma [30]. JMJD6 overexpression could induces EMT, and greatly enhances tumor metastasis in neuroblastoma and breast cancer [9, 27]. These results suggest that JMJD6 is involved in promoting cell transformation, tumor progression, and metastasis. In addition, considering the potential value of JMJD6 in cancer therapy, an inhibitor SKLB325 has been designed based on the crystal structure of the JmjC domain of JMJD6, which has shown significant anti-tumor effects in ovarian cancer [31]. It could be speculated that targeting JMJD6 might be a potential strategy for developing new therapies for ESCC.

Gene enrichment analysis showed that proteins co-expressed with JMJD6 were strongly associated with tumor progression, such as BRD4, MED12, CDK9, KDM8, TP53 and etc. Kaplan-Meier Plotter website was used to explore the prognostic value of the top 20 genes similar to JMJD6, and 5 genes were found to be statistically significant, all of which were positively correlated

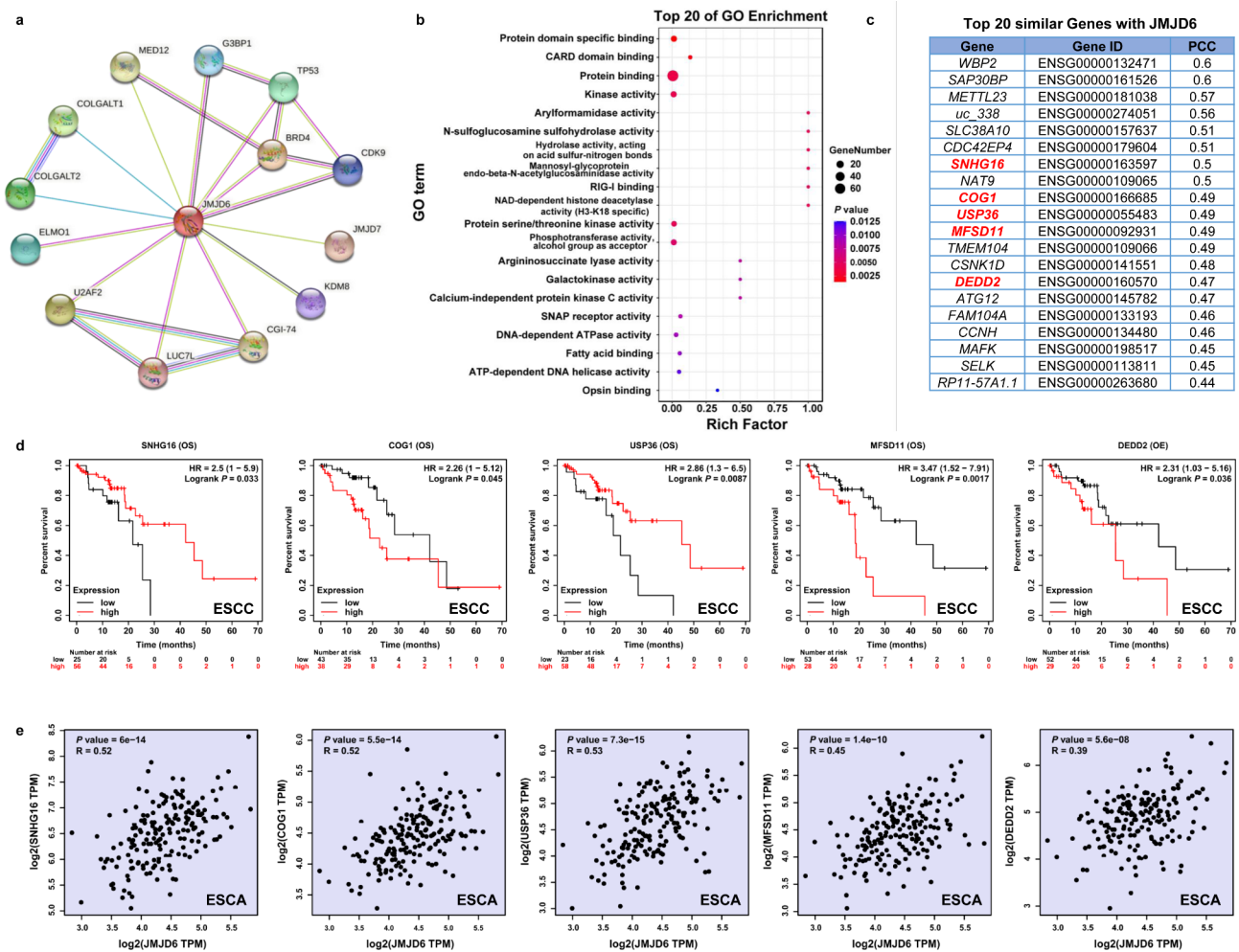


Fig. 3 JMJD6-related gene enrichment analysis. **(a)** Co-expression network of 14 proteins co-expressed with JMJD6 obtained by the STRING tool. **(b)** Gene Ontology analysis of the genes similar to JMJD6 obtained by the GEPIA2. **(c)** The top 20 genes similar to JMJD6 obtained by the GEPIA2. **(d)** Kaplan-Meier survival analysis of COG1, MFSD11, DEDD2, SNHG16 and USP36 expression from the Kaplan-Meier plotter database. **(e)** Correlation analysis between JMJD6 and COG1, MFSD11, DEDD2, SNHG16 and USP36 conducted by GEPIA2 across ESCA. Abbreviations: COG1, Conserved oligomeric Golgi 1; DEDD2, Death Effector Domain Containing 2; ESCA, esophageal carcinoma; ESCC, Esophageal squamous cell carcinoma; JMJD6, Jumonji domain-containing protein 6; MFSD11, Major facilitator superfamily domain 11; SNHG16, Small nucleolar RNA host gene 16

with JMJD6 expression in ESCA. However, high expression of COG1, MFSD11, DEDD2 while low expression of SNHG16 and USP36 was related to the poor prognosis of ESCC. We speculated that this result might be related to the fact that ESCC and EAC are two subtypes of esophageal cancer with different epidemiology and pathogenesis, different molecular profiles, as well as different therapeutic targets. In addition, we found that the genes similar to JMJD6 might act as oncogenes in the carcinogenic progression and are highly correlated with poor prognosis in multiple cancers.

COG1, a heteromeric subunit of the COG complex, plays a crucial role in endosomal to Golgi transport, retrograde transport of Golgi vesicles, and Golgi homeostasis [32]. Various studies have demonstrated that COG complex plays an important role in tumor metastasis by

regulating protein glycosylation and is associated with tumor grading and prognosis [33, 34]. Furthermore, COG complex dysfunction affects the dissociation of glycosyltransferases from anterograde cargo molecules and interferes with normal protein glycosylation [35]. Although the role of COG complex in tumor has not been reported, abnormal expression of COG complex can affect tumor invasion and metastasis by regulating protein glycosylation [32]. MFSD11 gene encoded protein contained major facilitator superfamily domain, this superfamily was a diverse group of secondary transporters [36]. However, the Uniprot website states that MFSD11 is related to the unc-93 family and not to the major promoter superfamily. There are less studies on MFSD11, thus the mechanism and role of MFSD11 are still unclear. A spearman correlation analysis showed

that MFSD11 was most significantly related to survival in ovarian cancer [37].

DEDD2, is a DEDD homolog, including an NH₂-terminal region with a DED domain, and a COOH-terminal region with some DNA-binding proteins [38]. DEDD2 is involved in regulating the degradation of intermediate filaments during apoptosis [39]. In addition, along with DEDD, DEDD2 is a strong inducer of death receptor-induced apoptosis [38]. It has been involved in a variety of malignancies. Studies have found that DEDD2 could downregulate miR-301 to help luteolin inhibit prostate cancer cell proliferation and induce apoptosis [40]. Furthermore, DEDD2 contributes to apoptosis of non-small cell lung cancer and triple-negative breast cancer cells [41]. Bioinformatics analysis found that the expression of *COG1*, *MFSD11* and *DEDD2* was positively correlated with JMJD6 and the poor prognosis of ESCC, which further proved the role of JMJD6 in the biological behavior of ESCC and deserved further exploration.

Conclusions

In a nutshell, we aimed to investigate JMJD6 in the occurrence and prognosis of ESCC through clinical data and bioinformatics analysis in public datasets. The obtained data indicated that high JMJD6 expression was significantly related with poor OS in ESCC patients. In addition, the analysis between JMJD6 expression and clinicopathological features as well as IHC staining of ESCC patients indicated that JMJD6 was a promising prognostic biomarker for ESCC patients. However, the mechanism of JMJD6 as a tumor-associated protein in ESCC is not sufficient, which need more researches to be further elucidated.

List of abbreviations

COG1	Conserved oligomeric Golgi 1
DEDD2	Death Effector Domain Containing 2
ESCA	Esophageal carcinoma
ESCC	Esophageal squamous cell carcinoma
GEPIA2	Gene Expression Profiling Interactive Analysis version 2
IHC	Immunohistochemical
JMJD6	Jumonji domain-containing protein 6
MFSD11	Major facilitator superfamily domain 11
OS	Overall survival
SNHG16	Small nucleolar RNA host gene 16

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s12885-023-11171-z>.

Supplementary Material 1

Acknowledgements

Not applicable.

Authors' contributions

Material preparation, data collection and analysis were performed by Liu H, Jiang M, and Ma F. Study conception and design was performed by Xu L, Yan X, and Jiang T. Figure design and table making were performed by Qin J,

Zhou X. The first draft of the manuscript was written by Xu L, Liu H. All authors commented on previous versions of the manuscript, read and approved the final manuscript.

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Data Availability

The dataset generated or analyzed during the current study are available in the GEPIA2 (<http://gepia2.cancer-pku.cn/>), Kaplan-Meier plotter database (<https://kmplot.com/>), and STRING tool (<https://string-db.org/>). The other data used during the current study are available from the corresponding author.

Declarations

Ethics approval and consent to participate

This study was performed in line with the principles of the Declaration of Helsinki. Ethics approval was granted by the Ethics Committee of Tangdu Hospital (No. TDLL-202110-02). This study was a retrospective study, and informed consent to participate was waived by Institution Review Board of Tangdu Hospital, Air Force Medical University.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

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References

- Li R, Zeng L, Zhao H, Deng J, Pan L, Zhang S, Wu G, Ye Y, Zhang J, Su J, et al. ATXN2-mediated translation of TNFR1 promotes esophageal squamous cell carcinoma via m(6)A-dependent manner. *Mol therapy: J Am Soc Gene Therapy*. 2022;30(3):1089–103.
- Abnet CC, Arnold M, Wei WQ. Epidemiology of esophageal squamous cell carcinoma. *Gastroenterology*. 2018;154(2):360–73.
- Watanabe M, Toh Y, Ishihara R, Kono K, Matsubara H, Miyazaki T, Morita M, Murakami K, Muro K, Numasaki H, et al. Comprehensive registry of esophageal cancer in Japan, 2015. *Esophagus: official journal of the Japan Esophageal Society*. 2023;20(1):1–28.
- Klose RJ, Kallin EM, Zhang Y. JmjC-domain-containing proteins and histone demethylation. *Nat Rev Genet*. 2006;7(9):715–27.
- Chang B, Chen Y, Zhao Y, Bruick RK. JMJD6 is a histone arginine demethylase. *Sci (New York NY)*. 2007;318(5849):444–7.
- Liu Y, Long YH, Wang SQ, Zhang YY, Li YF, Mi JS, Yu CH, Li DY, Zhang JH, Zhang XJ. JMJD6 regulates histone H2A.X phosphorylation and promotes autophagy in triple-negative breast cancer cells via a novel tyrosine kinase activity. *Oncogene*. 2019;38(7):980–97.
- Yang J, Chen S, Yang Y, Ma X, Shao B, Yang S, Wei Y, Wei X. Jumonji domain-containing protein 6 protein and its role in cancer. *Cell Prolif*. 2020;53(2):e12747.
- Shen X, De Geyter C, Zhang H, Huang G. Regulatory role of JMJD6 in placental development. *Expert Rev Mol Med*. 2022;24:e34.

9. Lee YF, Miller LD, Chan XB, Black MA, Pang B, Ong CW, Salto-Tellez M, Liu ET, Desai KV. JMJD6 is a driver of cellular proliferation and motility and a marker of poor prognosis in breast cancer. *Breast cancer research: BCR*. 2012;14(3):R85.
10. Wen C, Xu M, Mo C, Cheng Z, Guo Q, Zhu X. JMJD6 exerts function in neuropathic pain by regulating NF- κ B following peripheral nerve injury in rats. *Int J Mol Med*. 2018;42(1):633–42.
11. Lawrence P, Rai D, Conderino JS, Uddowla S, Rieder E. Role of Jumonji C-domain containing protein 6 (JMJD6) in infectivity of foot-and-mouth disease virus. *Virology*. 2016;492:38–52.
12. Ganesan M, Tikhanovich I, Vangimalla SS, Dagur RS, Wang W, Poluektova LI, Sun Y, Mercer DF, Tuma D, Weinman SA, et al. Demethylase JMJD6 as a New Regulator of Interferon Signaling: Effects of HCV and ethanol metabolism. *Cell Mol Gastroenterol Hepatol*. 2018;5(2):101–12.
13. Lee CR, Lee SH, Rigas NK, Kim RH, Kang MK, Park NH, Shin KH. Elevated expression of JMJD6 is associated with oral carcinogenesis and maintains cancer stemness properties. *Carcinogenesis*. 2016;37(2):119–28.
14. Ge Y, Liu BL, Cui JP, Li SQ. Livin promotes colon cancer progression by regulation of H2A.X(Y39ph) via JMJD6. *Life Sci*. 2019;234:116788.
15. Zhang Z, Yang Y, Zhang X. MiR-770 inhibits tumorigenesis and EMT by targeting JMJD6 and regulating WNT/ β -catenin pathway in non-small cell lung cancer. *Life Sci*. 2017;188:163–71.
16. Wan J, Liu H, Yang L, Ma L, Liu J, Ming L. JMJD6 promotes hepatocellular carcinoma carcinogenesis by targeting CDK4. *Int J Cancer*. 2019;144(10):2489–500.
17. Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. *Nucleic Acids Res*. 2017;45(W1):W98–w102.
18. Liao X, Bu Y, Xu Z, Jia F, Chang F, Liang J, Jia Q, Lv Y. WISP1 predicts clinical prognosis and is Associated with Tumor Purity, Immunocyte Infiltration, and macrophage M2 polarization in Pan-Cancer. *Front Genet*. 2020;11:502.
19. Wang W, Tang YA, Xiao Q, Lee WC, Cheng B, Niu Z, Oguz G, Feng M, Lee PL, Li B, et al. Stromal induction of BRD4 phosphorylation results in chromatin remodeling and BET inhibitor resistance in Colorectal Cancer. *Nat Commun*. 2021;12(1):4441.
20. Siraj AK, Masoodi T, Bu R, Pratheeshkumar P, Al-Sanea N, Ashari LH, Abduljabbar A, Alhomoud S, Al-Dayel F, Alkuraya FS, et al. MED12 is recurrently mutated in Middle Eastern colorectal cancer. *Gut*. 2018;67(4):663–71.
21. Cidado J, Boiko S, Proia T, Ferguson D, Criscione SW, San Martin M, Pop-Damkov P, Su N, Roamio Franklin VN, Sekhar Reddy Chilamakuri C, et al. AZD4573 is a highly selective CDK9 inhibitor that suppresses MCL-1 and induces apoptosis in Hematologic Cancer cells. *Clin cancer research: official J Am Association Cancer Res*. 2020;26(4):922–34.
22. Wang HJ, Pochampalli M, Wang LY, Zou JX, Li PS, Hsu SC, Wang BJ, Huang SH, Yang P, Yang JC, et al. KDM8/JMJD5 as a dual coactivator of AR and PKM2 integrates AR/EZH2 network and tumor metabolism in CRPC. *Oncogene*. 2019;38(1):17–32.
23. Lacroix M, Riscal R, Arena G, Linares LK, Le Cam L. Metabolic functions of the tumor suppressor p53: implications in normal physiology, metabolic disorders, and cancer. *Mol metabolism*. 2020;33:2–22.
24. Sooraj D, Sun C, Doan A, Garama DJ, Dannappel MV, Zhu D, Chua HK, Mahara S, Wan Hassan WA, Tay YK, et al. MED12 and BRD4 cooperate to sustain cancer growth upon loss of mediator kinase. *Mol Cell*. 2022;82(1):123–139e127.
25. Kosai-Fujimoto Y, Itoh S, Yugawa K, Fukuhara T, Okuzaki D, Toshima T, Harada N, Oda Y, Yoshizumi T, Mori M. Impact of JMJD6 on intrahepatic cholangiocarcinoma. *Mol Clin Oncol*. 2022;17(2):131.
26. Zhao J, Adams A, Roberts B, O'Neil M, Vittal A, Schmitt T, Kumer S, Cox J, Li Z, Weinman SA, et al. Protein arginine methyl transferase 1- and Jumonji C domain-containing protein 6-dependent arginine methylation regulate hepatocyte nuclear factor 4 alpha expression and hepatocyte proliferation in mice. *Hepatology (Baltimore MD)*. 2018;67(3):1109–26.
27. Wong M, Sun Y, Xi Z. JMJD6 is a tumorigenic factor and therapeutic target in neuroblastoma. *Nat Commun*. 2019;10(1):3319.
28. Wan J, Xu W, Zhan J, Ma J, Li X, Xie Y, Wang J, Zhu WG, Luo J, Zhang H. PCAF-mediated acetylation of transcriptional factor HOXB9 suppresses lung adenocarcinoma progression by targeting oncogenic protein JMJD6. *Nucleic Acids Res*. 2016;44(22):10662–75.
29. Grypari IM, Pappa I, Papastergiou T, Zolota V, Bravou V, Melachrinou M, Megalooikonomou V, Tzelepi V. Elucidating the role of PRMTs in prostate cancer using open access databases and a patient cohort dataset. *Histol Histopathol*. 2023;38(3):287–302.
30. Liu X, Si W, Liu X, He L, Ren J, Yang Z, Yang J, Li W, Liu S, Pei F, et al. JMJD6 promotes melanoma carcinogenesis through regulation of the alternative splicing of PAK1, a key MAPK signaling component. *Mol Cancer*. 2017;16(1):175.
31. Zheng H, Tie Y, Fang Z. Jumonji domain-containing 6 (JMJD6) identified as a potential therapeutic target in ovarian cancer. *Signal Transduct Target therapy*. 2019;4:24.
32. Zhang Y, Lai H, Tang B. Abnormal Expression and Prognosis Value of COG Complex Members in Kidney Renal Clear Cell Carcinoma (KIRC). *Disease markers* 2021, 2021:4570235.
33. Christiansen MN, Chik J, Lee L, Anugraham M, Abrahams JL, Packer NH. Cell surface protein glycosylation in cancer. *Proteomics*. 2014;14(4–5):525–46.
34. Oliveira-Ferrer L, Legler K, Milde-Langosch K. Role of protein glycosylation in cancer metastasis. *Sem Cancer Biol*. 2017;44:141–52.
35. Shestakova A, Zolov S, Lupashin V. COG complex-mediated recycling of golgi glycosyltransferases is essential for normal protein glycosylation. *Traffic*. 2006;7(2):191–204.
36. Gao Y, Wu N, Wang S, Yang X, Wang X, Xu B. Concurrent mutations associated with trastuzumab-resistance revealed by single cell sequencing. *Breast Cancer Res Treat*. 2021;187(3):613–24.
37. Yao S, Yuan C, Shi Y, Qi Y. Alternative splicing: a New Therapeutic Target for Ovarian Cancer. *Technol Cancer Res Treat*. 2022;21:15330338211067911.
38. Alcivar A, Hu S, Tang J, Yang X. DEDD and DEDD2 associate with caspase-8/10 and signal cell death. *Oncogene*. 2003;22(2):291–7.
39. Lee JC, Schickling O, Stegh AH, Oshima RG, Dinsdale D, Cohen GM, Peter ME. DEDD regulates degradation of intermediate filaments during apoptosis. *J Cell Biol*. 2002;158(6):1051–66.
40. Han K, Meng W, Zhang JJ, Zhou Y, Wang YL, Su Y, Lin SC, Gan ZH, Sun YN, Min DL. Luteolin inhibited proliferation and induced apoptosis of prostate cancer cells through miR-301. *Oncotargets and therapy*. 2016;9:3085–94.
41. Roth W, Stenner-Liewen F, Pawlowski K, Godzik A, Reed JC. Identification and characterization of DEDD2, a death effector domain-containing protein. *J Biol Chem*. 2002;277(9):7501–8.

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