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Haplotype analysis of TP53 polymorphisms, Arg72Pro and Ins16, in BRCA1 and BRCA2 mutation carriers of French Canadian descent

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Abstract

Background: The TP53 polymorphisms Arg72Pro (Ex4+199 G>C) and Ins16 (IVS3+24 ins16) have been proposed to modify risk of breast cancer associated with germline BRCA1 and BRCA2 mutations. Allele frequencies of these polymorphisms were investigated to determine if they modify risk in BRCA mutation carriers in breast cancer cases drawn from French Canadian cancer families, a population shown to exhibit strong founder effects.

Methods: The frequencies of the TP53 alleles, genotypes and haplotypes of 157 index breast cancer cases comprised of 42 BRCAI mutation carriers, 57 BRCA2 mutation carriers, and 58 BRCA mutation-negative cases, where each case was drawn from independently ascertained families were compared. The effect of TP53 variants on the age of diagnosis was also investigated for these groups. The TP53 polymorphisms were also investigated in 112 women of French Canadian descent with no personal history of cancer.

Results: The BRCA mutation-positive groups had the highest frequency of homozygous carriers of the 72Pro allele compared with mutation-negative group. The TP53 polymorphisms exhibited linkage disequilibrium (p < 0.001), where the 72Arg and Ins16minus alleles occurred in strong disequilibrium. The highest frequency of carriers of Ins16minus-72Arg haplotype occurred in the BRCA mutation-negative groups. The BRCA1 mutation carriers homozygous for the 72Pro allele had the youngest ages of diagnosis of breast cancer. However none of these observations were statistically significant. In contrast, the BRCA2 mutation carriers homozygous for the 72Pro allele had a significantly older age of diagnosis of breast cancer (p = 0.018). Moreover, in this group, the mean age of diagnosis of breast cancer in carriers of the Ins16minus-72Arg haplotype was significantly younger than that of the individuals who did not this carry this haplotype (p = 0.009).

Conclusion: We observed no significant association of breast cancer risk with TP53 genetic variants based on BRCAI/2 mutation carrier status. Although the small sample size did not permit analysis of all possible haplotypes, we observed that BRCA2 mutation carriers harboring the InsI6minus-72Arg haplotype had a significantly younger mean age of diagnosis of breast cancer. These observations suggest that investigations in a larger French Canadian sample are warranted to further elucidate the effects of TP53 variants on age of diagnosis of breast cancer among BRCAI and BRCA2 mutation carriers.

Background

Approximately 40% of French Canadian breast and/or ovarian cancer families have been shown to harbor germline mutations in the BRCA1 and BRCA2 cancer susceptibility genes [1-3]. At least five specific mutations in these genes have been found to recur in cancer families of French Canadian descent [2-5] and this has been attributed to common founders [1,3,6-9]. Germline mutations in BRCA1 and BRCA2 confer a high lifetime risk for breast and/or ovarian cancer, and early studies of familial cancer cases suggested that these risks may be as high as 80% [10,11]. However, lower estimates of lifetime risk for breast cancer of 66% in BRCA1 carriers and 45% in BRCA2 carriers were reported in subsequent populationbased studies of pooled data [12,13]. Although various host factors may influence or modify risk, such as parity [14], genetic factors have also been proposed as modifiers of risk, such as genetic variants of HRAS1 [15], the androgen receptor (AR) [16], the 5'UTR of RAD51 [17], and repeat length polymorphisms in AIB1 [18], not all of which have been replicated or substantiated in subsequent studies [19-21].

Genetic variants of TP53 have received attention as possible modifiers of cancer risk due to the critical role of p53 in cell cycle control, DNA repair, and apoptosis, and possible interaction with BRCA1 and BRCA2 [22-24]. Germline mutations in TP53 also confer significantly increased risk for hereditary breast cancer in the context of the Li Fraumeni syndrome and Li Fraumeni-like syndrome families, however the overall contribution is less than that observed for BRCA1 and BRCA2, as was also shown in a recent study of French Canadian breast and/or ovarian cancer families [25]. The Arg72Pro (Ex4+199 G>C) and Ins16 (IVS3+24 ins16) TP53 polymorphisms have been extensively studied as putative breast cancer susceptibility variants with inconsistent results [26-42]. These variants have been shown to affect the in vitro apoptotic activity of p53 [43-45]. For example, the chemotherapeutic response was less favorable in ovarian cancer cases retaining the TP53 72Pro variant which possibly accounts for the overall poorer prognosis following treatment of such cases [46]. A significantly increased familial breast cancer risk for carriers of the Ins16 variant has also been reported |42|.

Evidence is also emerging that the 72Pro and Ins16 TP53 polymorphisms may modify risk in carriers of BRCA1 or BRCA2 mutations [45,47]. The 72Pro allele has been found associated with a younger age of diagnosis of breast cancer in BRCA1 mutation carriers [47]. In a recent study of Spanish breast and/or ovarian cancer families, the haplotype lacking the Ins16 allele (referred to as "Ins16minus") and containing the 72Pro allele was associated with a younger age of diagnosis of breast cancer in BRCA2 mutation carriers than other comparative groups based on BRCA mutation status [45]. Recently we reported the frequency of TP53 polymorphisms, including 72Pro and Ins16 alleles in a study of the contribution of germline TP53 mutations in French Canadian breast and/or ovarian cancer families tested negative for BRCA1 and BRCA2 mutations [25]. However, the frequency of these polymorphisms and their effect on breast cancer risk in this founder population was not determined. Therefore, the aim of the present study was to investigate the contribution of these polymorphisms in a selected series of breast cancer cases with known BRCA1 and BRCA2 mutation status that were each drawn from independently ascertained breast and breast/ovarian cancer families of French Canadian descent. We report the frequencies of these alleles in BRCA1 and BRCA2 mutation-positive cases, and compare these frequencies with mutation-negative familial breast cancer cases, as well as female French Canadian controls with no personal history of cancer. The haplotype frequencies are also reported. In addition, we investigate the influence of TP53 alleles on the age of diagnosis of breast cancer.

Methods

Breast cancer cases and controls

Genotype analyses were performed on 157 breast cancer cases where each case was drawn from an independently ascertained cancer family in order to reduce bias due to familial relationships. Each family had at least three confirmed cases of female breast cancer (diagnosed ≤ 65 years of age), epithelial ovarian cancer, or male breast cancer as described previously [1-3,25]. The affected individuals in each family were first-, second- or third-degree relatives (occurring within the same lineage) to the index case that was selected for TP53 genotype analysis. Index cases reported grandparental French Canadian ancestry from

Quebec, Canada. The index cases from 42 families were BRCA1 mutation-positive (age range at initial diagnosis of 27 to 65 years; 43.7 years mean age of diagnosis), from 57 families were BRCA2 mutation-positive (age range at initial diagnosis of 26 to 65 years; 42.7 years mean age of diagnosis), and 58 cases were mutation-negative (age range at initial diagnosis of 30 to 65 years; 47.2 years mean age of diagnosis) based on commercial sequencing service (Myriad Genetics®, Salt Lake City, UT). Within this group of index breast cancer cases there were nine breast cancer cases with primary cancers of the breast and ovary, of which six individuals were BRCA1 mutation-positive, two individuals were BRCA2 mutation-positive, and one case was mutation-negative. The families were ascertained through the Service de Médecine Génique, Centre Hospitalier de l'Université de Montréal (CHUM) and the Hereditary Cancer Clinics of McGill University in Montreal. The controls were comprised of 112 females participants with no personal history of breast or ovarian cancer ascertained from the French Canadian population of Quebec.

The clinical samples (pheripheral blood lymphocytes), and personal and family history were attained from the study participants at the Centre de recherche du Centre hospitalier de l'Université de Montréal – Hôpital Hotel-Dieu and Institut du cancer de Montréal with signed informed consent as part of the tissue and clinical banking activities of the Banque de tissus et de données of the Réseau de recherche sur le cancer of the Fonds de la Recherche en Santé du Québec (FRSQ). The study was granted ethical approval from the Research Ethics Boards of the participating research institutes.

Genotype and haplotype analyses

The Arg72Pro polymorphism refers to the Ex4+199 G>C variant rs1042522 at nucleotide position 12139, which results in the nonsynonymous amino acid substitution of an arginine (72Arg) amino acid at codon 72 with a proline (72Pro) amino acid. The Ins16 polymorphism refers to the IVS3+24insACCTGGAGGGCTGGGG, the intronic variant rs17878362 at nucleotide position 11951. For simplicity genotypes lacking the Ins16 allele are referred

to as "Ins16minus". The nucleotide position is based on the TP53 reference sequence X54156. The genotyping assays were performed on DNA extracted from peripheral blood leukoctyes. The Arg72Pro and Ins16 polymorphisms were assayed as described previously [25]. Haplotypes were determined as described in Osorio A. et al., 2006 [45].

Statistical analyses

Allele, genotype and haplotype frequency distributions were compared by Monte-Carlo χ^2 analyses (Statistical Product and Service Solution Package, SPSS, Chicago, IL). Hardy-Weinberg equilibrium (HWE) for each polymorphism was tested by Monte-Carlo Markov Chain [48]. Linkage disequilibrium analysis and D' estimation were performed using the Arlequin v2.0 software package. A one-way ANOVA test was used to compare age related effects (onset of breast cancer). Kruskal-Wallis test was used where appropriate.

Results

Allele and Genotype frequencies in cases and controls

The genotype and allele frequencies of the TP53 polymorphisms from 112 controls and 157 index breast cancer cases each selected from an independently ascertained cancer family was determined. The controls had more individuals that were homozygous for the 72Pro allele than that of the cases, and this was also reflected in a higher 72Pro allele frequency (Table 1). In contrast, the cases had more individuals that were homozygous for the Ins16 allele than that the controls (Table 2). These findings are also reflected in the allele frequencies for these polymorphisms. However, the differences in genotype frequencies are not significant (Tables 1 and 2).

Haplotype analysis revealed that the TP53 polymorphisms are in linkage disequilibrium (D' = 0.80 - 0.93, for all test groups, p < 0.001), where the 72Arg and Ins16minus alleles occurred in strong disequilibrium. All possible double haplotypes were observed in the cases (Table 3), whereas there were no examples of Ins16-72Arg, Ins16-72Pro or Ins16-72Arg, Ins16minus-72Pro

Table I: Frequency of Ex4+199 G>C (72Pro) allele

Group	Sampl number e	Genotypic frequencies (%)			P values			Allele frequ	HWE	
		GG (Arg, Arg)	GC (Arg, Pro)	CC (Pro, Pro)	All cases	BRCA2+	BRCA-	G	С	P value
Controls	112	57 (50.9)	46 (41.1)	9 (8.0)	0.84			160 (71.4)	64 (28.6)	ı
All cases	157	80 (51.0)	67 (42.7)	10 (6.3)				227 (72.3)	87 (27.7)	0.55
BRCAI+	42	20 (47.6)	18 (42.9)	4 (9.5)		0.86	0.47	58 (69.0)	26 (31.0)	1
BRCA2+	57	29 (50.9)	24 (42.1)	4 (7.0)			0.78	82 (71.9)	32 (28.1)	1
BRCA-	58	31 (53.4)	25 (43.1)	2 (3.4)				87 (75.0)	29 (25.0)	0.47
BRCAI/2+	99	49 (49.5)	42 (42.5)	8 (8.0)			0.57	140 (70.7)	58 (29.3)	1

Table 2: Frequency of IVS3+24 ins16 (+) allele

Group		Genotypic frequencies (%)			P values			Allele frequ	HWE	
	Sample number		-+	+ +	All cases	BRCA2+	BRCA-	-	+	P value
Controls	112	79 (70.5)	32 (28.6)	I (0.9)	0.65			190 (84.8)	34 (15.2)	0.46
All cases	157	102 (65.0)	53 (33.8)	2 (1.2)				257 (81.8)	57 (18.2)	0.11
BRCAI+	42	26 (61.9)	16 (38.1)	Ò		0.48	0.83	68 (81.0)	16 (19.0)	0.31
BRCA2+	57	38 (66.7)	17 (29.8)	2 (3.5)			0.46	93 (81.6)	21 (18.4)	I
BRCA-	58	38 (65.5)	20 (34.5)	Ò				96 (82.8)	20 (17.2)	0.19
BRCAI/2+	99	64 (64.7)	33 (33.3)	2 (2.0)			0.76	161 (81.3)	37 (18.7)	0.18

double haplotypes in the controls. However, there was only one example of each of these double haplotypes in the cases. Moreover, the distribution of the double haplotypes in the cases did not differ significantly from that of the controls. The paucity of some double haplotypes did not permit an analysis of all possible double haplotypes. However, notable is that the distribution of the Ins16minus-72Arg double haplotype in the cases (145 of 157, 92.4%) and controls (103 of 112, 91.7%) were similar (OR = 1.06 (95% CI 0.44 - 2.54), p = 1.

Genotype and Allele Frequencies of Hereditary and Familial Cases of Breast Cancer

The genotype frequencies of the TP53 polymorphisms in each of the three breast cancer cases groups based on BRCA mutation status were determined. Each of the BRCA1 and BRCA2 mutation-positive groups had the highest frequency of homozygous carriers of the 72Pro allele when compared with that of the mutation-negative group (Table 1). Homozygous carriers of the Ins16 allele were only observed in BRCA2 mutation-positive carriers (Table 2). However, neither of these differences nor the distribution of all genotypes were significantly different in all pair-wise comparisons of the three groups, or when the BRCA mutation-positive groups were combined in the analyses for each polymorphism.

The distribution of all possible double haplotypes in the cases is shown in Table 3. None of the breast cancer case groups had individuals with all possible double haploytpes. The highest frequency of carriers of the Ins16minus-72Arg haplotype, which is in strong disequilibrium, was within the mutation-negative group. The BRCA mutationnegative group had the highest frequency of carriers of the Ins16minus-72Arg haplotype (56 of 58, 96.6%). However, this observation was not significantly different from that observed in either of the BRCA1 mutation-positive (38 of 42, 90.5%) and BRCA2 mutation-positive (51 of 57, 89.5%) groups (p = 0.29).

Age effects

Overall, the BRCA mutation-negative cases had a significantly older mean age of diagnosis of breast cancer than each of the BRCA1 and BRCA2 mutation-positive groups (p = 0.02), as has been observed in previous studies of French Canadian breast cancer families [2]. The differences in age of diagnosis of breast cancer in the BRCA1 and BRCA2 mutation-positive groups did not permit the combined analysis of these groups versus mutation-negative group. We therefore investigated the ages of diagnoses of breast cancer within each group separately with respect to each TP53 polymorphism (Figure 1). Within the BRCA1 mutation-positive group, homozygous 72Pro carriers had the youngest ages of diagnoses of breast cancer than that of the homozygous 72Arg (p = 0.31) and heter-

Table 3: Frequency of double haplotypes

	Double haplotype frequencies (%)											P value			
Group	Sample number	-G -G	-G +G	-G -C	-G +C	+G -C	+G +C	-c -c	-C +C	+C +C	All cases	BRCA2 +	BRCA-		
Control s	112	55 (49.1)	2 (1.8)	21 (18.8)	25 (22.3)	0	0	2 (1.8)	6 (5.4)	I (I.0)	0.98				
All cases	157	74 (47.1)	5 (3.2)	26 (16.6)	40 (25.5)	I (0.6)	I (0.6)	2 (1.2)	7 (4.5)	I (0.6)					
BRCAI +	42	18 (42.9)	2 (4.8)	7 (16.7)	11 (26.2)	0	0	I (2.4)	3 (7.1)	0		0.94	0.61		
BRCA2 +	57	26 (45.6)	2 (3.5)	12 (21.1)	11 (19.3)	I (I.8)	I (I.8)	0	3 (5.3)	I (I.8)			0.34		
BRCA-	58	30 (51.7)	1 (1.7)	7 (12.1)	18 (31.0)	0	0	1 (1.7)	1 (1.7)	0					
BRCAI/ 2+	99	44 (44.4)	4 (4.0)	19 (19.2)	22 (22.2)	I (I.0)	I (I.0)	I (I.0)	6 (6.1)	I (I.0)			0.61		

ozygous (p = 0.36) carriers, although these observations were not significant. Within the BRCA2 mutation-positive group, homozygous 72Pro carriers had a significantly older age of diagnosis of breast cancer compared to the mean age of diagnosis of either of the homozygous 72Arg (p = 0.041) and heterozygous (p = 0.018) carriers. Within the BRCA2 mutation-positive group, homozygous Ins16 carriers had a significantly older age of diagnosis of breast cancer compared to the mean age of diagnosis of either of the homozygous Ins16minus (p = 0.042) or heterozygous (p = 0.046) carriers.

The low frequency of all double haplotypes did not permit an investigation of each possible combination with respect to age of diagnosis of breast cancer (Figure 2). However, we were able to analyze the combined data for carriers of the Ins16minus and 72Arg alleles as these alleles were in strong linkage disequilibrium. Within the BRCA2 mutation-positive group, carriers of the Ins16minus-72Arg haplotype had a significantly younger mean age of diagnosis of breast cancer (p = 0.009). Indeed the mean age of diagnosis of breast cancer at 41.7 years of the 51 Ins16minus-72Arg carriers was significantly younger than that of the six cases that did not carry this haplotype.

Discussion

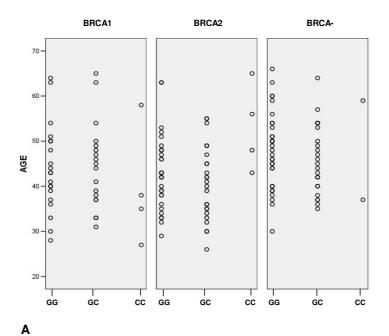
While there was an increased frequency of homozygous carriers of 72Pro alleles in BRCA1 and BRCA2 mutation carriers, and an increased frequency of homozygous carriers of Ins16 alleles in BRCA2 mutation carriers in familial breast cancer cases of French Canadian descent, the findings were not significant. The distribution of genotype frequencies was not significantly different in the controls or cancer cases in this study. Moreover, the distribution of genotypes and alleles of the unaffected controls and cases were comparable and not significantly different than that of European (or Caucasian) population as reported in the Single Nucleotide Polymorphism Database [49], a population that is historically or ancestrally linked with the French Canadians of Quebec [7]. These results suggest that there are no significant differences in the genotype frequencies in breast cancer cases regardless of the BRCA1 or BRCA2 mutation status. In independent reports of TP53 allele frequencies in cancer cases of BRCA1/2-mutation carriers, the relationships among carriers is not always apparent. In our study, in order to reduce bias due to the possibility of close familial relationships, we have purposely drawn our cases from independently ascertained families where (to our knowledge) the family members are not known to be directly related to each other [1-3]. However, our study size may be limited and further analysis of larger sample groups is warranted given the apparent increased frequency of homozygous carriers

of rare alleles for both genotypes in BRCA1/2 mutation carriers when compared with mutation-negative cases.

Although there were differences in the ages of diagnosis of breast cancer in homozygous carriers of the 72Pro allele in BRCA1 and BRCA2 mutation positive groups, the mean age of diagnoses were not significantly different. BRCA2 mutation carriers homozygous for the 72Pro allele had an older mean age of diagnosis of breast cancer compared with breast cancer cases homozygous for the 72Arg allele and heterozygous carriers of this TP53 polymorphism, but overall the difference was not statistically significant. Although our studies were limited in size and are not significant, the younger mean age of diagnosis of breast cancer in BRCA1 mutation carriers homozygous for the 72Pro allele is consistent with independent reports suggesting that this allele may modify penetrance of BRCA1 carriers [47]. The independent report of the effect on age of diagnosis of breast cancer of the Ins16minus-72Pro haplotype in BRCA2 mutation carriers [45] was not observed in our study. However, we observed that among the BRCA2 mutation-positive cases, carriers of the Ins16minus-72Arg haplotype had a significantly younger mean age of diagnosis of breast cancer compared to the other individuals within this group. However, haplotype analysis suggested that the two polymorphic loci were in strong linkage disequilibrium in our samples. Although the extent of the linkage disequilibrium for the TP53 has not been properly investigated in this population, haplotype analysis of BRCA1 and BRCA2 loci of carriers of recurrent mutations have shown it could extend beyond at least one centri-Morgan [1,3,6]. This observation was not surprising given the strong founder effects observed in the French Canadian population and the fact that the present day population are descendents of an estimated 8,500 settlers who colonized the present day St. Lawrence valley ("Nouvelle France") in recent history between 1608 and 1759 [7,8]. Hence, the young age of this founder population may also explain the relative paucity of individuals with the Ins16-72Arg haplotype in our sample groups.

Conclusion

The analysis of TP53 alleles, 72Pro and Ins16, in French Canadians suggest that they do not significantly modify familial breast cancer risk. However our analyses may be affected by sample size and strong linkage disequilibrium observed in this population for the tested alleles. While additional breast cancer cases drawn from within each cancer families could have been included in our analyses as has been done in previous studies [25], this may further bias results due to the founder effects and linkage disequilibrium observed for the tested alleles in this population. However, the difference in homozygous allele frequencies of rare genotypes in this population warrants further investigation with a larger sample.



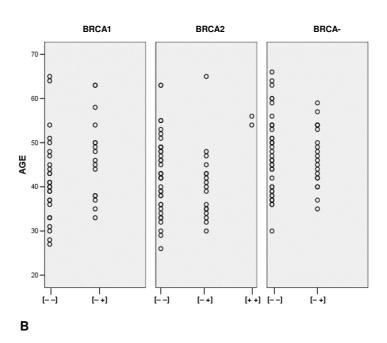


Figure 1Scatter plots of the age at diagnoses of breast cancer based on BRCA mutation status and genotypes of each TP53 polymorphism.

Panel A contains the age distribution of Arg72pro polymorphism based on BRCA mutation status for homozygous carriers of 72Arg (GG) and 72Pro (CC) genotypes or heterozygous carriers (GC). For the BRCA1 mutation-positive group, the G,G carriers (n = 20) had a mean age ± st. err. (standard error) = 43.9 ± 2.1; the GC carriers (n = 18) had mean age ± st. err. = 44.4 ± 2.3; and the CC carriers (n = 4) had a mean age ± st. err. = 39.5 ± 6.6. For the BRCA2 mutation-positive group, the GG carriers (n = 29) had a mean age ± st. err. = 43 ± 1.6; the GC carriers (n = 24) had a mean age ± st. err. = 40.5 ± 1.7; and the CC carrier (n = 4) had a mean age ± st. err. = 53 ± 4.8. For the BRCA mutation-negative group, the GG carriers (n = 31) had a mean age ± st. err. = 48.1 ± 1.5; the GC carriers (n = 25) had a mean age ± st. err. = 46.0 ± 1.5; and the CC carriers (n = 2) had a mean age ± st. err. = 48 ± 11. Panel B contains the age distribution of lns16 polymorphism based on BRCA mutation status for homozygous carriers lns16minus [- -] and ins16 [+ +] genotypes or heterozygous carriers [- +]. For the BRCA1 mutation-positive group, the [- -] carriers (n = 26) had a mean age ± st. err. = 41.7 ± 1.9; the [- +] carriers (n = 38) had a mean age ± st. err. = 43 ± 1.4; the [- +] carriers (n = 17) had a mean age ± st. err. = 40.5 ± 2.0; and the [+ +] carriers (n = 20) had a mean age ± st. err. = 46.9 ± 1.5; and there were no homozygous [+ +] carriers (n = 38) had a mean age ± st. err. = 46.9 ± 1.5; and there were no homozygous [+ +] carriers (n = 38) had a mean age ± st. err. = 46.9 ± 1.5; and there were no homozygous [+ +] carriers (n = 38) had a mean age ± st. err. = 46.9 ± 1.5; and there were no homozygous [+ +] carriers (n = 38) had a mean age ± st. err. = 46.9 ± 1.5; and there were no homozygous [+ +] carriers (n = 38) had a mean age ± st. err. = 47.3 ± 1.5; the [- +] carriers (n = 20) had a m

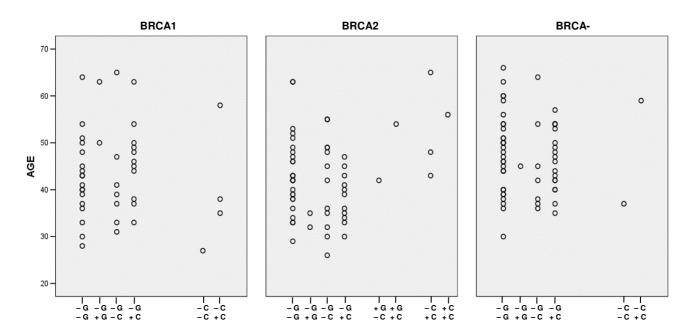


Figure 2 Scatter plots of the ages at diagnoses of breast cancer based on BRCA mutation status and the double haplotypes observed for the TP53 polymorphisms. Shown are the distributions of ages of diagnosis based on BRCA mutation status and TP53 polymorphism double haplotypes based on 72Arg [G], 72Pro [C], $\ln s \cdot 16 = 1$ and $\ln s \cdot 16$ minus [-] alleles. For the BRCA1 mutation-positive groups, the [-G; -G] carriers (n = 18) had a mean age \pm st. err. = 42.6 \pm 2.1; the [-G; +G] carriers (n = 2) had a mean age \pm st. err. = 56.5 \pm 6.5; the [-G; -C] carriers (n = 7) had a mean age \pm st. err. = 41.9 \pm 4.3; the [-G; +C] carriers (n = 11) mean age \pm st. err. = 46.1 \pm 2.5; the [-C; -C] carrier (n = 1) had a age = 27; and the [-C; +C] carriers (n = 3) had a mean age \pm st. err. = 43.7 \pm 7.2. For the BRCA2 mutation-positive carriers, the [-G; -G] carriers (n = 26) had a mean age \pm st. err. = 43.5 \pm 1.5; the [-G; +C] carriers (n = 12) had a mean age \pm st. err. = 41.8 \pm 2.8; the [-G; +C] carriers (n = 11) had a age = 54; the [-G; +C] carriers (n = 3) had a mean age \pm st. err. = 52.0 \pm 6.6; and the [+C; +C] carrier (n = 1) had a age = 56. For the BRCA mutation-negative group, the [-G; -G] carriers (n = 30) had a mean age \pm st. err. = 48.2 \pm 1.6; the [-G; +G] carrier (n = 1) had an age = 45; the [-G; -C] carriers (n = 7) had a mean age \pm st. err. = 48.2 \pm 1.6; the [-G; +G] carrier (n = 1) had an age = 37; and the [-C; +C] carrier (n = 1) had an age = 59.

Competing interests

The author(s) declare that they have no competing interest.

Authors' contributions

LC carried out the molecular genetic and statistical analyses, participated in study design and drafted the manuscript. SLA participated in the molecular genetic analyses. CM, PG, A-M M-M and DP provided the clinical information of cancer families, BRCA1 and BRCA2 mutation status, and DNA samples from cancer families, and PNT conceived of the study, and participated in its design, statistical analyses and coordination, and drafted the manuscript. All authors read and approved the final manuscript.

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References

- Tonin PN, Mes-Masson AM, Futreal PA, Morgan K, Mahon M, Foulkes WD, Cole DE, Provencher D, Ghadirian P, Narod SA: Founder BRCA1 and BRCA2 mutations in French Canadian breast and ovarian cancer families. Am J Hum Genet 1998, 63:1341-1351.
- Oros KK, Ghadirian P, Greenwood CM, Perret C, Shen Z, Paredes Y, Arcand SL, Mes-Masson AM, Narod SA, Foulkes WD, Provencher D, Tonin PN: Significant proportion of breast and/or ovarian cancer families of French Canadian descent harbor I of 5 BRCAI and BRCA2 mutations. Int J Cancer 2004, 112:411-419.
- Oros KK, Leblanc G, Arcand SL, Shen Z, Perret C, Mes-Masson AM, Foulkes WD, Ghadirian P, Provencher D, Tonin PN: Haplotype analysis suggest common founders in carriers of the recurrent BRCA2 mutation, 3398delAAAAG, in French Canadian hereditary breast and/ovarian cancer families. BMC Med Genet 2006, 7:23.
- Antoniou AC, Durocher F, Smith P, Simard J, Easton DF: BRCAI and BRCA2 mutation predictions using the BOADICEA and BRCAPRO models and penetrance estimation in high-risk French-Canadian families. Breast Cancer Res 2006, 8:R3.
- Simard J, Dumont M, Moisan AM, Gaborieau V, Malouin H, Durocher F, Chiquette J, Plante M, Avard D, Bessette P, Brousseau C, Dorval M,

- Godard B, Houde L, Joly Y, Lajoie MA, Leblanc G, Lepine J, Lesperance B, Vezina H, Parboosingh J, Pichette R, Provencher L, Rheaume J, Sinnett D, Samson C, Simard JC, Tranchant M, Voyer P, Easton D, Tavtigian SV, Knoppers BM, Laframboise R, Bridge P, Goldgar D: Evaluation of BRCA1 and BRCA2 mutation prevalence, risk prediction models and a multistep testing approach in French-Canadian families with high risk of breast and ovarian cancer. J Med Genet 2007, 44:107-121.
- Manning AP, Ábelovich D, Ghadirian P, Lambert JA, Frappier D, Provencher D, Robidoux A, Peretz T, Narod SA, Mes-Masson AM, Foulkes WD, Wang T, Morgan K, Fujiwara TM, Tonin PN: Haplotype analysis of BRCA2 8765delAG mutation carriers in French Canadian and Yemenite Jewish hereditary breast cancer families. Hum Hered 2001, 52:116-120.
- Scriver CR: Human genetics: lessons from Quebec populations. Annu Rev Genomics Hum Genet 2001, 2:69-101.
- Laberge AM, Michaud J, Richter A, Lemyre E, Lambert M, Brais B, Mitchell GA: Population history and its impact on medical genetics in Quebec. Clin Genet 2005, 68:287-301.
- Vezina H, Durocher F, Dumont M, Houde L, Szabo C, Tranchant M, Chiquette J, Plante M, Laframboise R, Lepine J, Nevanlinna H, Stoppa-Lyonnet D, Goldgar D, Bridge P, Simard J: Molecular and genealogical characterization of the R1443X BRCA1 mutation in high-risk French-Canadian breast/ovarian cancer families. Hum Genet 2005, 117:119-132.
- Ford D, Easton DF, Stratton M, Narod S, Goldgar D, Devilee P, Bishop DT, Weber B, Lenoir G, Chang-Claude J, Sobol H, Teare MD, Struewing J, Arason A, Scherneck S, Peto J, Rebbeck TR, Tonin P, Neuhausen S, Barkardottir R, Eyfjord J, Lynch H, Ponder BA, Gayther SA, Zelada-Hedman M, .: Genetic heterogeneity and penetrance analysis of the BRCA1 and BRCA2 genes in breast cancer families. The Breast Cancer Linkage Consortium. Am J Hum Genet 1998, 62:676-689.
- Easton DF, Ford D, Bishop DT: Breast and ovarian cancer incidence in BRCA1-mutation carriers. Breast Cancer Linkage Consortium. Am J Hum Genet 1995, 56:265-271.
- Antoniou AC, Easton DF: Polygenic inheritance of breast cancer: Implications for design of association studies. Genet Epidemiol 2003, 25:190-202.
- Simchoni S, Friedman E, Kaufman B, Gershoni-Baruch R, Orr-Urtreger A, Kedar-Barnes I, Shiri-Sverdlov R, Dagan E, Tsabari S, Shohat M, Catane R, King MC, Lahad A, Levy-Lahad E: Familial clustering of site-specific cancer risks associated with BRCA1 and BRCA2 mutations in the Ashkenazi Jewish population. Proc Natl Acad Sci U S A 2006, 103:3770-3774.
- Narod SA: Modifiers of risk of hereditary breast cancer. Oncogene 2006, 25:5832-5836.
- 15. Phelan CM, Rebbeck TR, Weber BL, Devilee P, Ruttledge MH, Lynch HT, Lenoir GM, Stratton MR, Easton DF, Ponder BA, Cannon-Albright L, Larsson C, Goldgar DE, Narod SA: Ovarian cancer risk in BRCAI carriers is modified by the HRASI variable number of tandem repeat (VNTR) locus. Nat Genet 1996, 12:309-311.
- Rebbeck TR, Kantoff PW, Krithivas K, Neuhausen S, Blackwood MA, Godwin AK, Daly MB, Narod SA, Garber JE, Lynch HT, Weber BL, Brown M: Modification of BRCAI-associated breast cancer risk by the polymorphic androgen-receptor CAG repeat. Am J Hum Genet 1999, 64:1371-1377.
- Levy-Lahad E, Lahad A, Eisenberg S, Dagan E, Paperna T, Kasinetz L, Catane R, Kaufman B, Beller U, Renbaum P, Gershoni-Baruch R: A single nucleotide polymorphism in the RAD51 gene modifies cancer risk in BRCA2 but not BRCA1 carriers. Proc Natl Acad Sci U S A 2001, 98:3232-3236.
- Rebbeck TR, Wang Y, Kantoff PW, Krithivas K, Neuhausen SL, Godwin AK, Daly MB, Narod SA, Brunet JS, Vesprini D, Garber JE, Lynch HT, Weber BL, Brown M: Modification of B. Cancer Res 2001, 61:5420-5424.
- 19. Hughes DJ, Ginolhac SM, Coupier I, Barjhoux L, Gaborieau V, Bressac-de-Paillerets B, Chompret A, Bignon YJ, Uhrhammer N, Lasset C, Giraud S, Sobol H, Hardouin A, Berthet P, Peyrat JP, Fournier J, Nogues C, Lidereau R, Muller D, Fricker JP, Longy M, Toulas C, Guimbaud R, Yannoukakos D, Mazoyer S, Lynch HT, Lenoir GM, Goldgar DE, Stoppa-Lyonnet D, Sinilnikova OM: Breast cancer risk in BRCA1 and BRCA2 mutation carriers and polyglutamine repeat length in the AIB1 gene. Int J Cancer 2005, 117:230-233.

- Kadouri L, Kote-Jarai Z, Hubert A, Durocher F, Abeliovich D, Glaser B, Hamburger T, Eeles RA, Peretz T: A single-nucleotide polymorphism in the RAD51 gene modifies breast cancer risk in BRCA2 carriers, but not in BRCA1 carriers or noncarriers. Br J Cancer 2004, 90:2002-2005.
- Spurdle AB, Antoniou AC, Kelemen L, Holland H, Peock S, Cook MR, Smith PL, Greene MH, Simard J, Plourde M, Southey MC, Godwin AK, Beck J, Miron A, Daly MB, Santella RM, Hopper JL, John EM, Andrulis IL, Durocher F, Struewing JP, Easton DF, Chenevix-Trench G: The AIBI polyglutamine repeat does not modify breast cancer risk in BRCA1 and BRCA2 mutation carriers. Cancer Epidemiol Biomarkers Prev 2006, 15:76-79.
- Cheung AM, Elia A, Tsao MS, Done S, Wagner KU, Hennighausen L, Hakem R, Mak TW: Brca2 deficiency does not impair mammary epithelium development but promotes mammary adenocarcinoma formation in p53(+/-) mutant mice. Cancer Res 2004, 64:1959-1965.
- Jonkers J, Meuwissen R, van der GH, Peterse H, van V, Berns A: Synergistic tumor suppressor activity of BRCA2 and p53 in a conditional mouse model for breast cancer. Nat Genet 2001, 29:418-425.
- Ongusaha PP, Ouchi T, Kim KT, Nytko E, Kwak JC, Duda RB, Deng CX, Lee SW: BRCAI shifts p53-mediated cellular outcomes towards irreversible growth arrest. Oncogene 2003, 22:3749-3758.
- Arcand SL, Maugard CM, Ghadirian P, Robidoux A, Perret C, Zhang P, Fafard E, Mes-Masson AM, Foulkes WD, Provencher D, Narod SA, Tonin PN: Germline TP53 mutations in BRCA1 and BRCA2 mutation-negative French Canadian breast cancer families. Breast Cancer Res Treat 2007.
- Sjalander A, Birgander R, Hallmans G, Cajander S, Lenner P, Athlin L, Beckman G, Beckman L: p53 polymorphisms and haplotypes in breast cancer. Carcinogenesis 1996, 17:1313-1316.
- 27. Weston A, Godbold JH: Polymorphisms of H-ras-I and p53 in breast cancer and lung cancer: a meta-analysis. Environ Health Perspect 1997, 105 Suppl 4:919-926.
- Wang-Gohrke S, Rebbeck TR, Besenfelder W, Kreienberg R, Runnebaum IB: p53 germline polymorphisms are associated with an increased risk for breast cancer in German women. Anticancer Res 1998, 18:2095-2099.
- Papadakis EN, Dokianakis DN, Spandidos DA: p53 codon 72 polymorphism as a risk factor in the development of breast cancer. Mol Cell Biol Res Commun 2000, 3:389-392.
- Wang-Gohrke S, Becher H, Kreienberg R, Runnebaum IB, Chang-Claude J: Intron 3 16 bp duplication polymorphism of p53 is associated with an increased risk for breast cancer by the age of 50 years. *Pharmacogenetics* 2002, 12:269-272.
- Buyru N, Tigli H, Dalay N: P53 codon 72 polymorphism in breast cancer. Oncol Rep 2003, 10:711-714.
- Huang XE, Hamajima N, Katsuda N, Matsuo K, Hirose K, Mizutani M, Iwata H, Miura S, Xiang J, Tokudome S, Tajima K: Association of p53 codon Arg72Pro and p73 G4C14-to-A4T14 at exon 2 genetic polymorphisms with the risk of Japanese breast cancer. Breast Cancer 2003, 10:307-311.
- Suspitsin EN, Buslov KG, Grigoriev MY, Ishutkina JG, Ulibina JM, Gorodinskaya VM, Pozharisski KM, Berstein LM, Hanson KP, Togo AV, Imyanitov EN: Evidence against involvement of p53 polymorphism in breast cancer predisposition. Int J Cancer 2003, 103:431-433.
- Kalemi TG, Lambropoulos AF, Gueorguiev M, Chrisafi S, Papazisis KT, Kotsis A: The association of p53 mutations and p53 codon 72, Her 2 codon 655 and MTHFR C677T polymorphisms with breast cancer in Northern Greece. Cancer Lett 2005, 222:57-65.
- 35. Noma C, Miyoshi Y, Taguchi T, Tamaki Y, Noguchi S: Association of p53 genetic polymorphism (Arg72Pro) with estrogen receptor positive breast cancer risk in Japanese women. Cancer Lett 2004, 210:197-203.
- Ohayon T, Gershoni-Baruch R, Papa MZ, Distelman MT, Eisenberg BS, Friedman E: The R72P P53 mutation is associated with familial breast cancer in Jewish women. Br J Cancer 2005, 92:1144-1148.
- Tommiska J, Eerola H, Heinonen M, Salonen L, Kaare M, Tallila J, Ristimaki A, von SK, Aittomaki K, Heikkila P, Blomqvist C, Nevanlinna H:
 Breast cancer patients with p53 Pro72 homozygous genotype have a poorer survival. Clin Cancer Res 2005, 11:5098-5103.

- Damin AP, Frazzon AP, Damin DC, Roehe A, Hermes V, Zettler C, Alexandre CO: Evidence for an association of TP53 codon 72 polymorphism with breast cancer risk. Cancer Detect Prev 2006, 30:523-529.
- Ma H, Hu Z, Zhai X, Wang S, Wang X, Qin J, Chen W, Jin G, Liu J, Gao J, Wang X, Wei Q, Shen H: Joint effects of single nucleotide polymorphisms in P53BP1 and p53 on breast cancer risk in a Chinese population. Carcinogenesis 2006, 27:766-771.
- Chinese population. Carcinogenesis 2006, 27:766-771.
 40. Campbell IG, Eccles DM, Dunn B, Davis M, Leake V: p53 polymorphism in ovarian and breast cancer. Lancet 1996, 347:393-394.
- 41. Khaliq S, Hameed A, Khaliq T, Ayub Q, Qamar R, Mohyuddin A, Mazhar K, Qasim-Mehdi S: **P53 mutations, polymorphisms, and haplotypes in Pakistani ethnic groups and breast cancer patients.** Genet Test 2000, 4:23-29.
- 42. Wirtenberger M, Frank B, Hemminki K, Klaes R, Schmutzler RK, Wappenschmidt B, Meindl A, Kiechle M, Arnold N, Weber BH, Niederacher D, Bartram CR, Burwinkel B: Interaction of Werner and Bloom syndrome genes with p53 in familial breast cancer. Carcinogenesis 2006, 27:1655-1660.
- Dumont P, Leu JI, Della PAC III, George DL, Murphy M: The codon 72 polymorphic variants of p53 have markedly different apoptotic potential. Nat Genet 2003, 33:357-365.
- Thomas M, Kalita A, Labrecque S, Pim D, Banks L, Matlashewski G: Two polymorphic variants of wild-type p53 differ biochemically and biologically. Mol Cell Biol 1999, 19:1092-1100.
- Osorio A, Martinez-Delgado B, Pollan M, Cuadros M, Urioste M, Torrenteras C, Melchor L, Diez O, De La HM, Velasco E, Gonzalez-Sarmiento R, Caldes T, Alonso C, Benitez J: A haplotype containing the p53 polymorphisms Ins I 6bp and Arg72Pro modifies cancer risk in BRCA2 mutation carriers. Hum Mutat 2006, 27:242-248.
- Santos AM, Sousa H, Portela C, Pereira D, Pinto D, Catarino R, Rodrigues C, Araujo AP, Lopes C, Medeiros R: TP53 and P21 polymorphisms: response to cisplatinum/paclitaxel-based chemotherapy in ovarian cancer. Biochem Biophys Res Commun 2006, 340:256-262.
- 47. Martin AM, Kanetsky PA, Amirimani B, Colligon TA, Athanasiadis G, Shih HA, Gerrero MR, Calzone K, Rebbeck TR, Weber BL: Germline TP53 mutations in breast cancer families with multiple primary cancers: is TP53 a modifier of BRCA1? J Med Genet 2003, 40:e34.
- Guo SW, Thompson EA: A Monte Carlo method for combined segregation and linkage analysis. Am J Hum Genet 1992, 51:111-1126.
- Sherry ST, Ward MH, Kholodov M, Baker J, Phan L, Smigielski EM, Sirotkin K: dbSNP: the NCBI database of genetic variation. Nucleic Acids Res 2001, 29:308-311.

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