

RESEARCH ARTICLE

Open Access

# Matrix metalloproteinase 1 and circulating tumor cells in early breast cancer

Zuzana Cierna<sup>1</sup>, Michal Mego<sup>2,3,5\*</sup>, Pavol Janega<sup>1,6</sup>, Marian Karaba<sup>5</sup>, Gabriel Minarik<sup>4</sup>, Juraj Benca<sup>5</sup>, Tatiana Sedláčková<sup>4</sup>, Silvia Cingelova<sup>5</sup>, Paulina Gronosova<sup>7</sup>, Denisa Manasova<sup>2</sup>, Daniel Pindak<sup>5,8</sup>, Jozef Sufliarsky<sup>3,5</sup>, Ludovit Danihel<sup>1</sup>, James M Reuben<sup>9</sup> and Jozef Mardiak<sup>3,5</sup>

## Abstract

**Background:** Matrix metalloproteinases (MMPs) are involved in cancer invasion and metastasis. Circulating tumor cells (CTCs) play role in tumor dissemination and are an independent survival predictor in breast cancer (BC) patients. The aim of this study was to assess correlation between CTCs and tumor MMP1 in BC.

**Methods:** Study included 149 primary BC patients treated by surgery from March 2012 to March 2013. Peripheral blood mononuclear cells (PBMC) were depleted of hematopoietic cells using RosetteSep™ selection kit. RNA extracted from CD45-depleted PBMC was interrogated for expression of EMT (TWIST1, SNAIL1, SLUG, ZEB1) and epithelial (CK19) gene transcripts by qRT-PCR. Patient samples with higher epithelial and/or mesenchymal gene transcripts than those of healthy donors (n = 60) were considered as CTC positive. Expression of MMP1 in surgical specimens was evaluated by immunohistochemistry.

**Results:** CTCs were detected in 24.2% patients. CTCs exhibiting only epithelial markers were present in 8.7% patients, whereas CTCs with epithelial-mesenchymal transition (EMT) markers (CTC\_EMT) were observed in 13.4% of patients and CTCs co-expressing both markers were detected in 2.0% patients. Patients with CTC\_EMT in peripheral blood had significantly increased expression of MMP1 in tumor cells (p = 0.02) and tumor associated stroma (p = 0.05) than those of patients without CTC\_EMT. In multivariate analysis, CTC\_EMT and tumor grade were independently associated with MMP1 expression in cancer cells, while CTC\_EMT and Ki67 were independently associated with MMP1 expression in cancer associated stroma.

**Conclusion:** Our data suggest link between MMP1 and CTCs with EMT phenotype and support role of MMPs and EMT in tumor dissemination.

**Keywords:** Circulating tumor cells, Matrix-metalloproteinase 1, Primary breast cancer

## Background

Circulating tumor cells (CTCs) play an important role in the metastatic cascade, cancer dissemination and progression. Prognostic value of CTCs was showed by numerous trials for metastatic as well as primary breast cancers [1-4]. CTCs constitute a heterogeneous population of cells with different phenotypes, clinical and biological value [5].

Invasion is one of the first critical steps in the metastatic cascade that requires changes in cell-to-cell

adhesion and cell adhesion to the extracellular matrix (ECM). Invasion is often accompanied by downregulation of epithelial molecules on cell surface including E-cadherin, and upregulation of mesenchymal molecules like N-cadherin, which is closely associated with mesenchymal cells [6]. Invasion is further facilitated by proteolytic degradation of the ECM, which enables cancer cells to penetrate tissue boundaries. Degradation of the ECM is predominately mediated by matrix metalloproteinases (MMPs) and the urokinase plasminogen activator (uPa) system [7].

Matrix metalloproteinase 1 (MMP1) is a zinc-dependent endopeptidase with collagen-cleaving activity. MMP1 cleave extracellular matrix components and thus play an important role in tumor invasion [8]. MMP1 is produced

\* Correspondence: misomego@gmail.com

<sup>2</sup>Translational Research Unit, Comenius University, Bratislava, Slovakia

<sup>3</sup>2nd Department of Medical Oncology, Comenius University, Faculty of Medicine, National Cancer Institute, Klenova 1, 833 10 Bratislava, Slovak Republic

Full list of author information is available at the end of the article

by tumor cells as well as by tumor associated stroma. High MMP1 expression in tumor is associated with tumor evolution, poor prognosis and shortened survival in different types of tumors including breast cancer [9,10]. Many studies suggest that overexpression of MMPs is one of the key events leading to the breast cancer dissemination. Recently, it was shown, that MMPs induce epithelial to mesenchymal transition and thus increase the invasive potential of tumor cells [11,12].

In this translational study, we hypothesized, that MMP1 is involved in CTC release and CTCs are detected more often in breast cancer patients with high MMP1 expression in primary tumor or tumor associated stroma. Therefore, we examined expression of MMP1 on breast tumor tissue as well as tumor associated stroma and correlated them with CTCs in peripheral blood. We also correlated MMP1 expression with other patients' tumor characteristics.

## Methods

### Study patients

As a part of ongoing translational study (Protocol TRU-SK 002; Chair: M. Mego), treatment naive patients with stages I–III primary breast cancer (PBC) who were undergoing definitive surgery were included. From each patient we obtained peripheral blood for CTCs detection on the day of surgery and corresponding paraffin-embedded tumor tissue. Each patient was given a complete diagnostic evaluation to exclude the presence of distant metastasis. Patients with concurrent malignancy other than non-melanoma skin cancer in the previous 5 years were excluded as well. In all patients, data regarding age, tumor stage, histology, regional lymph node involvement, hormone receptor status, and HER2 status were also recorded. All patients agreed to participate in the study and signed informed consent according to the IRB-approved protocol.

The study was approved by the Institutional Review Board (IRB) of the National Cancer Institute of Slovakia and patients were enrolled between March 2012 and March 2013. Healthy donors (N = 60) were age-matched women without breast cancer who were recruited and consented according to the IRB-approved protocol. Healthy donors were recruited from staff working in cooperating institutions except women from departments directly involved in the study.

### Detection of CTC in peripheral blood

CTC were detected in peripheral blood by quantitative real-time polymerase chain reaction (qRT-PCR) based assay utilizing CD45 positive (CD45+) cells depletion for CTCs enrichment, as described previously [13].

### RNA extraction

Peripheral blood was subjected to CD45 depletion using RosetteSep™ kit (StemCell technologies) according to the manufacturer's instructions. CD45-depleted cells were mixed with TRIzolVR LS Reagent (Invitrogen Corporation, Carlsbad, CA) and stored at  $-80^{\circ}\text{C}$  until it was necessary to extract RNA according to the manufacturer's instructions. All RNA preparation and handling steps took place in a laminar flow hood, under RNase-free conditions. RNA concentration was determined by absorbance readings at 260 nm.

### Identification of gene transcripts in CD45-enriched subsets

Isolated RNA was subjected to quantitative RT-PCR (qRT-PCR) to detect EMT-inducing transcription factors (EMT-TF) gene transcripts (TWIST, SNAIL1, SLUG and ZEB1) and epithelial antigen (CK19). In brief, 2.5  $\mu\text{L}$  of cDNA were placed in 25  $\mu\text{L}$  of reaction volume containing 12.5  $\mu\text{L}$  of QuantiFast Probe RT-PCR Kit Master Mix, 0.25  $\mu\text{L}$  QuantiFast RT Mix, 8.5  $\mu\text{L}$  water and 1.25  $\mu\text{L}$  of primers. The following TaqMan assays were purchased from LifeTechnologies (USA): Twist1: Hs00361186\_m1; Snail1: Hs00195591\_m1; Slug: Hs00161904\_m1; Zeb1: Hs01566408\_m1; Gapdh Hs99999905\_m1; Ck19 Hs00761767\_s1. Amplicons or probes spanned intron–exon boundaries, with the exception of CK19. Amplification was performed on an Eppendorf Realplex Real-Time PCR system (Eppendorf, Germany) using the cycling program:  $95^{\circ}\text{C}$  for 10 min; 40 cycles of  $95^{\circ}\text{C}$  for 15 sec and  $60^{\circ}\text{C}$  for 60 sec. All samples were analyzed in triplicate. Calibrator samples were run with every plate to ensure consistency of the PCR. For all fluorescence-based RT-PCR, fluorescence was detected between 0 and 40 cycles for the control and marker genes in single-plex reactions, which allowed for the deduction of the cycles at threshold (Ct) value for each product. Expression of the genes of interest was calibrated against expression of the housekeeping gene, GAPDH. Target cDNA was quantified using the delta-Ct method with the formula:  $1 = 2^{\text{Ct}(\text{target-GAPDH})}$ .

### CTC definition

Patient samples with higher Ck19 gene transcripts than those of healthy donors were considered as epithelial CTCs positive (CTC\_EP), while patient samples with higher EMT-TF (Twist1, Snail1, Slug and Zeb1) gene transcripts than those of healthy donors were considered as CTC\_EMT positive.

The highest expression levels of the CK19 and EMT-inducing TF gene transcripts relative to that of Gapdh were  $3.4 \times 10^{-3}$ ,  $2.0 \times 10^{-4}$ ,  $1 \times 10^{-2}$  and  $2.2 \times 10^{-2}$  for Ck19, Twist1, Snail1 and Zeb1, while Slug transcripts were not detected in any of the samples from healthy donor.

These values were used as “cutoff” to determine CTCs positivity.

### **Tumor pathology**

Pathology review was conducted at the Department of Pathology, Faculty of Medicine, Comenius University, by pathologists (ZC, LD and PJ) associated with the study.

### **Diagnosis and tumor samples**

The study included tumor specimens from 149 patients. All specimens were classified according to the WHO Classification of 2004. The block containing the most representative part of the haematoxylin and eosin (H&E) stained tumor was identified by microscopy and subsequently used for IHC analysis.

### **Tissue microarray construction**

According to tumor histology, one or two representative tumor areas were identified on H&E stained sections. Sections were matched to their corresponding wax blocks (the donor blocks), and 3-mm diameter cores of the tumor were removed from donor blocks with the multi-purpose sampling tool Harris Uni-Core (Sigma-Aldrich, Steinheim, Germany) and inserted into the recipient master block. The recipient block was cut into 5- $\mu$ m sections, and the sections were transferred to coated slides.

### **Immunohistochemical (IHC) staining**

Slides were deparaffinised and rehydrated in phosphate buffered saline solution (10 mM, pH 7.2). The tissue epitopes were demasked using the automated water bath heating process in Dako PT Link (Dako, Glostrup, Denmark); the slides were incubated in pH 6.0 citrate retrieval buffer at 98°C for 20 minutes. The slides were subsequently incubated overnight at room temperature with the primary rabbit polyclonal antibody against MMP1 (LSBio, MMP-1, LS-B1229) diluted 1:40 in Dako REAL antibody diluent (Dako, Glostrup, Denmark) and immunostained using anti-mouse/anti-rabbit immuno-peroxidase polymer (EnVision FLEX/HRP, Dako, Glostrup, Denmark) for 30 minutes at room temperature, according to the manufacturer's instructions. For visualisation, the slides were reacted with diaminobenzidine substrate-chromogen solution (DAB, Dako, Glostrup, Denmark) for 5 minutes. Finally, the slides were counterstained with haematoxylin. As negative control, breast tissue was subjected to the same staining procedure without reaction with the primary antibody.

### **Immunohistochemical stain scoring**

Tumor cores were independently assessed by pathologists (ZC, LD and PJ) who were blinded to clinico-pathological data. In cases of disagreement, the result was reached by consensus. The result of the immunohistochemical

analyses was expressed by a weighted histoscore, evaluating both the percentage of positive cells (PP) and the staining intensity (SI) of the nuclei or cytoplasm. Briefly, the proportion of cells with nuclear staining was multiplied by the intensity of staining to provide a histoscore ranging from 0–300. The histoscore was calculated as follows: Score = (0  $\times$  percentage not stained) + (1  $\times$  percentage weakly stained) + (2  $\times$  percentage moderately stained) + (3  $\times$  percentage strongly stained) [14].

### **Statistical analysis**

Patient characteristics were tabulated. The patients' characteristics were summarized using the median (range) for continuous variables and frequency (percentage) for categorical variables. Normality of distribution was tested by the Kolmogorov-Smirnoff test. If normally distributed, sample means were tested by Student *t*-test or analysis of variance (ANOVA) with Bonferroni's or Tamhane's corrections, depending on homogeneity of variance. Non-parametric Mann-Whitney *U* or Kruskal-Wallis *H* test were used for non-normally distributed data. A multivariate linear regression model was built using a step-wise regression technique, regression coefficients remained in the model at significance level of 0,05. MMP1 expression levels were analyzed as continuous variables. Because the MMP expression levels were highly skewed, Box-Cox transformations [15] were used for the determination of *p*-values, with optimum  $\lambda = 0,19$  for MMP1 in tumor stroma and optimum  $\lambda = 0,16$  for MMP1 expression in tumor cells. All *p* values presented are two-sided, and associations were considered significant if the *p* value is less or equal to 0.05. Statistical analyses were performed using NCSS 2007 software (Hintze J, 2007, Kaysville, Utah, USA).

### **Results**

The study population consisted of 149 primary breast cancer patients with median age of 60 years (range: 31–83 years). Patients' characteristics are shown in Table 1. The majority of patients had hormone receptor positive (87.2%), node negative (61.1%) tumors; 22 (14.8%) patients had HER-2/neu amplified tumors.

### **CTC detection**

To determine overexpression of the EMT-TF gene transcripts and Ck19 in PBC patients, we compared the expression levels in patient samples with those of HDs. Relative to the highest levels of Snail and Zeb1 transcripts detected in HD samples, none of the patient samples overexpressed these gene transcripts. Among the patient samples, Twist1, Slug, and Ck19 transcripts were

**Table 1 Patients characteristics**

Variable	N	%
All	149	100.0
<b>T-stage</b>		
1	105	70.5
> 1	44	29.5
<b>N-stage</b>		
0	91	61.1
> 1	58	38.9
<b>Grade</b>		
1 and 2	95	63.8
3	54	36.2
<b>Histology</b>		
IDC	128	85.9
Other	21	14.1
<b>Hormone receptor status</b>		
Negative	19	12.8
Positive	130	87.2
<b>HER2 status</b>		
Negative	127	85.2
Amplified	22	14.8
<b>Ki 67 (cut-off 20%)</b>		
Low	92	61.7
High	57	38.3
<b>Lymphovascular invasion</b>		
Present	35	23.5
Absent	114	76.5
<b>Baseline CTC</b>		
<b>CTC Epithelial</b>		
Negative	133	89.3
Positive	16	10.7
<b>CTC EMT</b>		
Negative	126	84.6
Positive	23	15.4
<b>CTC Any</b>		
Negative	113	75.8
Positive	36	24.2

overexpressed in 3 (2.0%), 20 (13.6%) and 16 (10.7%) samples, respectively. Totally, CTCs were detected in 36 (24.2%) of patients. CTCs with only epithelial markers were present in peripheral blood of 13 (8.7%) patients; CTCs with EMT phenotype were present in 20 (13.4%) of patients, while in 3 (2.0%) of patients CTCs exhibited both epithelial and mesenchymal markers. In one patient sample, there was overlap in overexpression of EMT-TF gene transcripts (Slug and Twist1).

### Correlation between MMP1 expression and CTC and patients'/tumor characteristics

MMP1 expression at least 1+ and higher was detected in 104 (69.8%) of samples in breast tumor cells and in 120 (80.5%) samples of tumor associated stroma ( $p = 0.04$ ) (Figure 1). Mean  $\pm$  SEM (standard error of mean) for MMP1 expression histoscore in tumor associated stroma was significantly higher compared to breast tumor cells ( $32.8 \pm 3.4$  vs.  $49.1 \pm 5.1$ ,  $p = 0.05$ ). Expression of MMP1 in relation to CTCs and various clinicopathological characteristics is shown in Table 2.

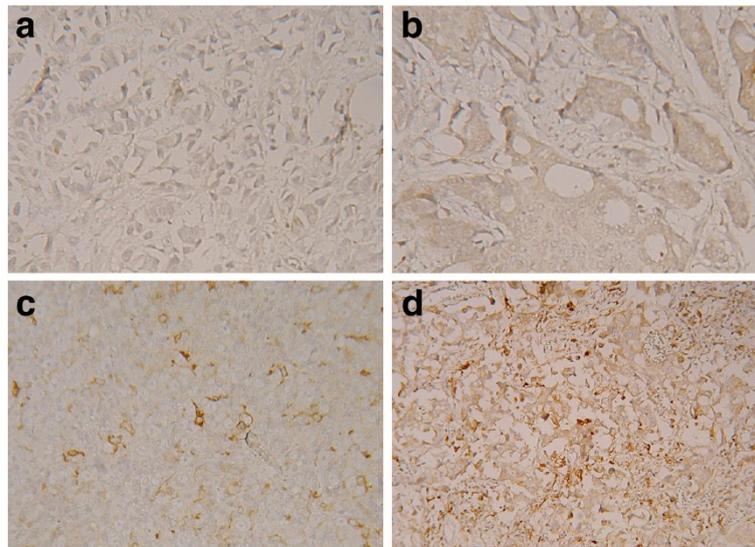
We observed an association between CTC\_EMT and expression of MMP1 in breast cancer cells as well as in cancer associated stroma (Table 2), and found no correlation between CTC\_EP and MMP1 expression. MMP1 expression in breast cancer cells was associated with high tumor grade, and increased proliferation (high Ki67), while MMP1 expression in tumor associated stroma, besides high tumor grade and increased proliferation, correlated with invasive ductal histology (Table 2). There was no association between expression of MMP1 and ER/PR status, HER2/neu amplification or axillary lymph node status (Table 2).

In multivariate analysis CTC\_EMT ( $p = 0.03$ ) and tumor grade ( $p = 0.01$ ) were independently associated with MMP1 expression in breast tumor cells, while Ki-67 ( $p = 0.0001$ ) and CTC\_EMT ( $p = 0.04$ ) were independently associated with MMP1 expression in tumor associated stroma (Tables 3 and 4).

### Discussion

In this translational study, we showed association between CTC with EMT phenotype (CTC\_EMT) and MMP1 expression in primary breast cancer. This association was observed for MMP1 expression in cancer cells as well as in cancer associated stroma. MMP1 expression was increased in primary tumors with poor prognostic features such as high grade tumors with increased proliferation (Ki 67 > 20%) which is consistent with previous observations [9]. We also found association between ductal carcinoma histology and MMP1 expression as described previously [16]; however, in our study, this association was statistically significant only for tumor associated stroma.

Breast cancer represents highly complex tissue composed of cancer cells and stromal cell compartments containing different types of mesenchymal cells. MMPs, in breast cancer, are produced by cancer cells as well as cancer associated stroma and are involved in cancer progression through multiple mechanisms. Degradation of ECM by MMPs facilitates movement of cancer cells through ECM. MMPs also disrupt cell-cell and cell-ECM adhesions that result in the release of individual tumor cells from epithelial sheets, and initiating signaling pathways



**Figure 1** MMP1 expression in primary breast tumors. Immunohistochemical reaction with anti-MMP1 monoclonal antibody. Original magnification  $\times 400$  visualisation with 3,3'-diaminobenzidine. **a)** staining intensity 0, **b)** staining intensity 1+, **c)** staining intensity 2+, **d)** staining intensity 3+.

that lead to widespread changes in gene expression patterns that are responsible for increased migration and invasion of breast cancer cells [7,11,17]. MMPs could also impact cancer cell behaviour due to their ability to cleave growth factors, cell surface receptors, cell adhesion molecules and chemokines [7,16]. Therefore, MMPs are promising therapeutic targets in breast cancer and several trials evaluating drugs that interfere with MMPs function are ongoing [reviewed by 11].

EMT is believed to play an important role in intravasation and the release of CTCs, and the expression of EMT-inducing TF gene transcripts in breast cancer has been associated with poor prognosis [18]. EMT has been previously linked with cancer stem cell properties [19] which have been associated with increased therapeutic resistance [20-22]. MMPs have been associated with EMT in cancer progression by several mechanisms [11]. Elevated levels of MMPs in the tumor microenvironment can directly induce EMT in epithelial cells. Cancer cells that undergo EMT could produce more MMPs further supporting cell invasion and metastasis and finally, EMT can generate activated stromal like cells that drive cancer progression via further MMPs production [11]. The most important of these is MMPs mediated activation of EMT that was seen in variety of epithelial tumors and has been best characterized in mammary epithelial cells [11]. Our observation thus further support association between MMPs and EMT in breast cancer.

EMT of tumor cells could produce stromal-like cells that could further facilitate tumor progression through the production of MMPs. Myofibroblasts are key components of cancer associated stroma in breast cancer

and these cells have important tumor promoting activity. Myofibroblasts can be derived through activation of stromal fibroblast or circulating fibrocytes; however, recent studies in mouse models have shown, that myofibroblasts can be derived from epithelial cells by EMT, as well [23-26]. In our study, we observed an association between CTC\_EMT and MMP1 expression in both cancer cells as well as in cancer associated stroma. Expression of MMP1 in the stromal compartment of breast carcinomas possibly represents two populations of cells: EMT transformed neoplastic cells and stromal fibroblastic cells that undergo activation of EMT induced TFs due to growth factors produced by the tumor [27]. However, we don't know exactly, if an observed association between CTC\_EMT and MMP1 in tumor stroma, was due to expression of MMP1 in stromal fibroblasts, or stromal like cancer cells that underwent EMT.

The flow of cancer cells may not be unidirectional. Experimental data suggest, that CTCs can be released from metastatic tumors and then rejoin the tumor of origin, a process termed 'self seeding' [28,29]. It seems, that self-seeding is a major driver of tumor progression in solid tumors and that CTCs are mediators of tumor self-seeding [28]. CTCs are an independent prognostic factor in breast cancer and probably represent surrogate marker for tumor self-seeding ability. In seminal work, Kim et al. observed that MMP1 is a mediator of CTCs infiltration into mammary tumors [28] and are involved in tumor self-seeding in an animal model. Our translational data are consistent with these observations and further support role of MMP1 in tumor self-seeding.

**Table 2 MMP1 expression in tumor cells and tumor stroma**

Variable	MMP1 expression histoscore in tumor cells					MMP1 expression histoscore in tumor associated stroma				
	N	Mean	SEM	Median	P-value	Mean	SEM	Median	P-value	
All	149	32.8	3.4	15	NA	49.1	5.1	20	NA	
<b>T-stage</b>										
1	105	30.6	4.0	14	0.28	49.7	6.1	20	0.76	
> 1	44	44.0	38.2	17		43.0	47.6	20		
<b>N-stage</b>										
0	91	32.1	4.4	14	1.00	44.7	6.5	20	0.29	
> 1	58	34.0	5.5	18		55.9	8.1	20		
<b>Grade</b>										
1 and 2	95	27.0	4.2	10	0.007	37.9	6.2	6	0.0006	
3	54	43.2	5.6	30		69.2	8.3	50		
<b>Histology</b>										
IDC	128	34.3	3.7	20	0.15	53.8	5.4	20	0.0022	
Other	21	23.6	9.0	4		20.8	13.3	1		
<b>Hormone receptor status</b>										
Negative	19	42.0	9.5	20	0.26	70.7	14.5	50	0.18	
Positive	130	31.5	3.6	13		46.1	5.4	20		
<b>HER2 status</b>										
Negative	127	30.4	3.7	10	0.18	45.0	5.5	20	0.29	
Amplified	22	47.1	8.8	25		72.5	13.1	40		
<b>Ki 67 (cut-off 20%)</b>										
Low	92	28.6	4.3	10	0.04	34.7	6.2	10	0.0006	
High	56	39.7	5.5	25		72.8	7.9	50		
<b>Lymphovascular invasion</b>										
Present	35	41.0	7.0	20	0.10	48.2	10.5	20	0.83	
Absent	114	30.3	3.9	11		49.4	5.8	20		
<b>Baseline CTC</b>										
<b>CTC Epithelial</b>										
Negative	133	32.9	3.6	16	0.35	48.6	5.4	20	0.50	
Positive	16	32.1	10.4	3		53.0	15.5	11		
<b>CTC EMT</b>										
Negative	126	30.2	3.7	10	0.02	45.7	5.5	20	0.05	
Positive	23	47.0	8.6	40		68.5	13.1	50		
<b>CTC Any</b>										
Negative	113	31.3	3.9	10	0.47	47.9	5.8	20	0.75	
Positive	36	37.5	6.9	20		53.0	10.5	20		

**Table 3 Multivariate logistic regression model for expression of MMP1 in tumor cells**

Parameter	T Statistic	P-value
Tumor grade (1 and 2 vs. 3)	2.59	0.0106
CTC_EMT present vs. absent	2.22	0.0277

**Table 4 Multivariate logistic regression model for expression of MMP1 in tumor associated stroma**

Parameter	T Statistic	P-value
Ki 67 (high vs. low)	3.94	0.0001
CTC_EMT present vs. absent	2.03	0.0444

The EMT is a dynamic and progressive process, and it can also be reversible. Therefore, it usually produces cells with a spectrum of intermediate phenotypic states. Cells can advance to differing extents through an EMT program, progressively acquiring mesenchymal features as they shed epithelial ones [5]. Cells that have entered an EMT program rarely shed all of their pre-existing epithelial features. In the context of carcinoma pathogenesis, neoplastic cells may reside in a state in which they coexpress newly acquired mesenchymal markers together with retained epithelial ones [13,30,31]. For these reasons our distinction of two CTCs subpopulation (CTC-EP, CTC-EMT) may represent some study limitations.

## Conclusion

In conclusion, in this prospective translational study, we for the first time showed association between CTC\_EMT and expression MMP1 in primary tumor tissue. We suppose that the therapeutical targeting of MMP1 could lead to decreased MMP1-induced EMT and subsequently, to decrease of CTC\_EMT, with implications for tumor dissemination and treatment resistance. Future studies will be needed to identify expression of other proteins in tumor tissue associated with presence of CTCs in the peripheral blood. These proteins could represent surrogate markers for biologically more aggressive disease and could represent potentially new therapeutic targets to inhibit metastatic process.

## Ethics approval

The Institutional Review Board of the National Cancer Institute, Bratislava, Slovakia.

## Abbreviations

ANOVA: Analysis of variance; CD45+: CD45 positive; CTCs: Circulating tumor cells; ECM: Extracellular matrix; EMT: Epithelial-to-mesenchymal transition; EMT-TF: EMT-inducing transcription factors; ER: Estrogen receptor; HD: Healthy donors; H&E: Hematoxylin and eosin; IRB: Institutional Review Board; IRS: ImmunoReactive Score; MBC: Metastatic breast cancer; MMPs: Matrix metalloproteinases; PB: Peripheral blood; PBMC: Peripheral blood mononuclear cells; PR: Progesterone receptor; qRT-PCR: Quantitative real time polymerase chain reaction; SEM: Standard error of the mean; TF: Tissue factor.

## Competing interest

On behalf of all the authors I declare that there are no competing financial interests in relation to the work described in the manuscript.

## Authors' contribution

M-M, J-R, J-S and J-M participated in conception and design of this study. Tumor cores were independently assessed by pathologists (ZC, LD and PJ) who were blinded to clinicopathological data. S-C performed statistical analysis, G-M, T-S, D-M were involved in CTCs detection, M-K, J-B, D-P were involved in patients accrual and performed breast surgery. M-M and ZC drafted the article and all authors reviewed it critically for important intellectual content. All the authors participated in the acquisition, analysis and interpretation of data. All the authors gave their final approval of the version to be published.

## Authors' information

Cierna Z and Mego M share first authorship.

## Acknowledgements

We would like to acknowledge our collaborators from department of pathology: Gabriela Sieberova, Jan Macuch, Michal Majercik, Peter Jani and Pavel Babal. We would like to acknowledge Zlatica Pekova for administration support and Emilia Klinecova and Ludovit Gaspar for excellent technical assistance. This publication is the result of the implementation of project funded by the Slovak Grant Agency [VEGA 1/0724/11 to M.M and J.M.]. Grant agency had no role in study design; in the collection, analysis, and interpretation of data; in the writing of the manuscript; and in the decision to submit the manuscript for publication.

## Author details

<sup>1</sup>Department of Pathology, Comenius University, Bratislava, Slovakia. <sup>2</sup>Translational Research Unit, Comenius University, Bratislava, Slovakia. <sup>3</sup>2nd Department of Medical Oncology, Comenius University, Faculty of Medicine, National Cancer Institute, Klenova 1, 833 10 Bratislava, Slovak Republic. <sup>4</sup>Institute of Molecular Biomedicine, Faculty of Medicine, Comenius University, Bratislava, Slovakia. <sup>5</sup>National Cancer Institute, Bratislava, Slovakia. <sup>6</sup>Institute of Normal and Pathological Physiology, Bratislava, Slovakia. <sup>7</sup>Cancer Research Institute, Slovak Academy of Sciences, Bratislava, Slovakia. <sup>8</sup>Slovak Medical University, Bratislava, Slovakia. <sup>9</sup>Department of Hematopathology, The University of Texas MD Anderson Cancer Center, Houston, Texas, USA.

Received: 17 March 2014 Accepted: 25 June 2014

Published: 28 June 2014

## References

1. Cristofanilli M, Budd GT, Ellis MJ, Stopeck A, Matera J, Miller MC, Reuben JM, Doyle GV, Allard WJ, Terstappen LW, Hayes DF: **Circulating tumor cells, disease progression, and survival in metastatic breast cancer.** *N Engl J Med* 2004, **351**:781–791.
2. Lucci A, Hall CS, Lodhi AK, Bhattacharyya A, Anderson AE, Xiao L, Bedrosian I, Kuerer HM, Krishnamurthy S: **Circulating tumour cells in non-metastatic breast cancer: a prospective study.** *Lancet Oncol* 2012, **13**:688–695.
3. Zhang L, Riethdorf S, Wu G, Wang T, Yang K, Peng G, Liu J, Pantel K: **Meta-analysis of the prognostic value of circulating tumor cells in breast cancer.** *Clin Cancer Res* 2012, **18**:5701–5710.
4. Zhao S, Liu Y, Zhang Q, Li H, Zhang M, Ma W, Zhao W, Wang J, Yang M: **The prognostic role of circulating tumor cells (CTCs) detected by RT-PCR in breast cancer: a metaanalysis of published literature.** *Breast Cancer Res Tr* 2011, **130**:809–816.
5. Mego M, Mani SA, Cristofanilli M: **Molecular mechanisms of metastasis in breast cancer-clinical applications.** *Nat Rev Clin Oncol* 2010, **7**:693–701.
6. Yang J, Weinberg RA: **Epithelial mesenchymal transition: at the crossroads of development and tumor metastasis.** *Dev Cell* 2008, **14**:818–829.
7. Egeblad M, Werb Z: **New functions for the matrix metalloproteinases in cancer progression.** *Nat Rev Cancer* 2002, **2**:161–174.
8. Kessenbrock K, Plaks V, Werb Z: **Matrix metalloproteinases: regulators of the tumor microenvironment.** *Cell* 2010, **141**:52–67.
9. Boström P, Söderström M, Vahlberg T, Söderström KO, Roberts PJ, Carpén O, Hirsimäki P: **MMP-1 expression has an independent prognostic value in breast cancer.** *BMC Cancer* 2011, **11**:348.
10. Roy R, Yang J, Moses MA: **Matrix metalloproteinases as novel biomarkers and potential therapeutic targets in human cancer.** *J Clin Oncol* 2009, **27**:5287–5297.
11. Radisky ES, Radisky DC: **Matrix metalloproteinase-induced epithelial-mesenchymal transition in breast cancer.** *J Mammary Gland Biol Neoplasia* 2010, **15**:201–212.
12. Mannello F: **What does matrix metalloproteinase-1 expression in patients with breast cancer really tell us?** *BMC Med* 2011, **9**:95.
13. Mego M, Mani SA, Lee BN, Li C, Evans KW, Cohen EN, Gao H, Jackson SA, Giordano A, Hortobagyi GN, Cristofanilli M, Lucci A, Reuben JM: **Expression of epithelial-mesenchymal transition-inducing transcription factors in primary breast cancer: The effect of neoadjuvant therapy.** *Int J Cancer* 2012, **130**:808–816.
14. van Nes JG, de Kruijf EM, Putter H, Faratian D, Munro A, Campbell F, Smit VT, Liefers GJ, Kuppen PJ, van de Velde CJ, Bartlett JM: **Co-expression of SNAIL and TWIST determines prognosis in estrogen receptor-positive early breast cancer patients.** *Breast Cancer Res Treat* 2012, **133**:49–59.
15. Box GEP, Cox DR: **An analysis of transformations.** *J R Stat Soc B* 1964, **26**:211–252.

16. Del Casar JM, González-Reyes S, González LO, González JM, Junquera S, Bongera M, García MF, Andicochea A, Serra C, Vizoso FJ: **Expression of metalloproteases and their inhibitors in different histological types of breast cancer.** *J Cancer Res Clin Oncol* 2010, **136**:811–819.
17. Page-McCaw A, Ewald AJ, Werb Z: **Matrix metalloproteinases and the regulation of tissue remodelling.** *Nat Rev Mol Cell Biol* 2007, **8**:221–233.
18. Martin TA, Goyal A, Watkins G, Jiang WG: **Expression of the transcription factors snail, slug, and twist and their clinical significance in human breast cancer.** *Ann Surg Oncol* 2005, **12**:488–496.
19. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M, Campbell LL, Polyak K, Brisken C, Yang J, Weinberg RA: **The epithelial-mesenchymal transition generates cells with properties of stem cells.** *Cell* 2008, **133**:704–715.
20. Hollier BG, Evans K, Mani SA: **The epithelial-to-mesenchymal transition and cancer stem cells: a coalition against cancer therapies.** *J Mammary Gland Biol Neoplasia* 2009, **14**:29–43.
21. Li X, Lewis MT, Huang J, Gutierrez C, Osborne CK, Wu MF, Hilsenbeck SG, Pavlick A, Zhang X, Chamness GC, Wong H, Rosen J, Chang JC: **Intrinsic resistance of tumorigenic breast cancer cells to chemotherapy.** *J Natl Cancer Inst* 2008, **100**:672–679.
22. Woodward WA, Chen MS, Behbod F, Alfaro MP, Buchholz TA, Rosen JM: **WNT/betacatenin mediates radiation resistance of mouse mammary progenitor cells.** *Proc Natl Acad Sci U S A* 2007, **104**:618–623.
23. Petersen OW, Nielsen HL, Gudjonsson T, Villadsen R, Rank F, Niebuhr E, Bissell MJ, Rønnov-Jessen L: **Epithelial to mesenchymal transition in human breast cancer can provide a nonmalignant stroma.** *Am J Pathol* 2003, **162**:391–402.
24. Willis BC, duBois RM, Borok Z: **Epithelial origin of myofibroblasts during fibrosis in the lung.** *Proc Am Thorac Soc* 2006, **3**:377–382.
25. Li JH, Wang W, Huang XR, Oldfield M, Schmidt AM, Cooper ME, Lan HY: **Advanced glycation end products induce tubular epithelial-myofibroblast transition through the RAGE-ERK1/2 MAP kinase signaling pathway.** *Am J Pathol* 2004, **164**:1389–1397.
26. Moinfar F, Man YG, Arnould L, Bratthauer GL, Ratschek M, Tavassoli FA: **Concurrent and independent genetic alterations in the stromal and epithelial cells of mammary carcinoma: implications for tumorigenesis.** *Cancer Res* 2000, **60**:2562–2566.
27. Soini Y, Tuhkanen H, Sironen R, Virtanen I, Kataja V, Auvinen P, Mannermaa A, Kosma VM: **Transcription factors zeb1, twist and snai1 in breast carcinoma.** *BMC Cancer* 2011, **11**:73.
28. Kim MY, Oskarsson T, Acharyya S, Nguyen DX, Zhang XH, Norton L, Massagué J: **Tumor self-seeding by circulating cancer cells.** *Cell* 2009, **139**:1315–1326.
29. Norton L: **Cancer stem cells, self-seeding, and decremented exponential growth: theoretical and clinical implications.** *Breast Dis* 2008, **29**:27–36.
30. Yu M, Bardia A, Wittner BS, Stott SL, Smas ME, Ting DT, Isakoff SJ, Ciciliano JC, Wells MN, Shah AM, Concampnon KF, Donaldson MC, Sequist LV, Brachtel E, Sgroi D, Baselga J, Ramaswamy S, Toner M, Haber DA, Maheswaran S: **Circulating breast tumor cells exhibit dynamic changes in epithelial and mesenchymal composition.** *Science* 2013, **339**:580–584.
31. Tam WL, Weinberg RA: **The epigenetics of epithelial-mesenchymal plasticity in cancer.** *Nat Med* 2013, **19**:1438–1449.

doi:10.1186/1471-2407-14-472

**Cite this article as:** Cierna et al.: Matrix metalloproteinase 1 and circulating tumor cells in early breast cancer. *BMC Cancer* 2014 **14**:472.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

